

Characterization of Viral Gene Proteins Panleukopenia Local Isolate as a Vaccine Candidate

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ABSTRACT

The purpose of this study was to determine the genetic characteristics of local FPV isolates in Indonesia as a basis for determining the type of FPV to be used as a panleukopenia vaccine material in Indonesia. This research was conducted in March - September 2019. Rectal swab or stool samples were taken, performed PCR and Sequencing, data were analyzed using the BioEdit program, homology analysis and phylogenetic trees using MEGA software. The results of this study found FPV isolates using PCR. In addition, based on the alignment results, the matching results of the sample sequences V1 and V2, respectively, are the Felocell® 3 vaccine and Purevax RCP. Whereas FPV virus field isolates namely V3 - V8 samples showed some differences in sequences namely in sequences 5, 95, 150, 173, and 425. Differences in nucleotide bases in several strains, vaccines and field isolates also showed a clear genetic difference in phylogenetic trees. Alignment of DNA sequences obtained homogeneous in several strains of FPV according to the NCBI database, which is precisely in the VP1 protein, VP2 FPV. The results of sequencing from eight samples showed a match on eight strains of FPV virus in a database with identities ranging from 99.2% - 99.8%. The conclusion from the results of this study is that FPV isolates were obtained and could be used as vaccine candidates for FPV in Indonesia. In addition, further studies on experimental animals are needed.

Keywords: Characteristics, FPV, cats, local isolates, vaccines

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BACKGROUND

Feline Panleukopenia (FPL) caused by Feline Parvovirus or better known as Feline Panleukopenia Virus (FPV) is still a serious disease problem in cats both in Indonesia and in the world. Feline Panleukopenia (FPL) is an infectious disease that causes high mortality (> 90%) with nerve symptoms.¹ Handling this disease in a period of more than 40 years still uses vaccinations developed in the era of the 60s.² While FPV continues to adapt and evolve, so there will be an inability of the vaccine to protect cats from the virus attack. This is thought to be due to mutations in the FPV.³ There are still many cases of cats infected with FPV after vaccination in Indonesia, but the number of events is not recorded, besides that the vaccination failure factor is still based on the assumption that cats are in an unhealthy condition at the time of vaccination. This can occur because of the suspected genetic differences or incompatibility of vaccine virus strains with field viruses. Vaccine that does not protect will create problems and losses because FPV can survive in the environment for many years and will be disinfectant resistant.^{4,5}

Based on the facts above that to reduce losses due to post-vaccination FPV infection, research needs to be done by looking at and analyzing the structure of the nucleotide bases and amino acids making up the gene in the FPV so that it can determine the genetic characteristics of local FPV isolates in Indonesia as a basis for determining the type of FPV to be used as a panleukopenia vaccine material in Indonesia.

RESEARCH METHODS

Time and Place of Research

This research was conducted in March - September 2019. Rectal swab or stool samples were taken in East Java. The

study was conducted at the Laboratory of Biosciences, Universitas Brawijaya Malang for primary ordering, the ADD (Animal Diagnostic Disease) Laboratory of the Faculty of Veterinary Medicine, Universitas Brawijaya for conducting research.

Rectal Swab Cat Sampling

Rectal swab sampling is performed after the handling procedure is performed. The location of the sample is taken in the rectum of the cat. Samples were taken using a sterile cotton swab and then swabed on the cat's rectum. Samples that have been taken are inserted into a viral transport medium (VTM) tube and labeled according to the individual's name. Samples that have been in the VTM tube are then stored in the freezer. Samples that have been frozen in the freezer are put into the ice box.

Polymerase Chain Reaction

DNA samples from cats infected with the panleukopenia virus were amplified using the PCR method. The pairs of primers used are forward primers (FPV_F) 5'- GCT TAC GCT TCT CTT CT-3 GCT 'and reverse (FPV_R) 5'- GCA TCA ACC AAT GAC CAA GG-3'. Amplification was started by mixing 3 µL DNA, 1.5 µL forward primer 10pmol, 1.5 µL reverse primer 10pmol, 15 µL PCR mix and 9 µL ddH₂O into a 200 µL microtube (PCR tube). The amplification step starts from predenaturing 94°C for four minutes, denaturing 94°C for 30 seconds, then annealing at 55°C for 30 seconds. Extension at 72°C for 1 minute and post extension at 72°C for 7 minutes. The process will repeat for 35 cycles.

DNA sequencing

PCR product sequencing from two directions is done by using the primary FPV_F and FPV_R to see the amplified sequences using the Sanger method. The PCR product shipped is a PCR product that has not been purified with an amount of 40 µL and the concentration of each primer

is 10 pmol. Sequencing results in the form of a graph stating the content of adenine, thymine, guanine, and cytosine contained in DNA fragments that have been labeled by ddNTPs.

Data analysis

The sequencing data in the form of ABI file was edited manually using the BioEdit program, homology analysis and phylogenetic tree using MEGA software.

RESULTS AND DISCUSSION

FPV is a non-zoonotic infectious disease that attacks cats in all ages and has a high mortality rate of 75%. The highest mortality and morbidity occur in cats aged one to twelve months. In the case of acute Panleukopenia the mortality ranges from 25-90% and 100% in silver infection. FPV can cause systemic infections and the most common enteric disease in kittens. The severity of FPV infection depends on age, immunity status, and the presence of secondary infection. Disease can be in the form of sub-clinical

infections and per-acute syndromes that can cause sudden death.⁶ There are still many cases of cats infected with FPV after vaccination in Indonesia, but the number of events is not recorded, besides that the vaccination failure factor is still based on the assumption that cats are in an unhealthy condition at the time of vaccination. This can occur because of the suspected genetic differences or incompatibility of vaccine virus strains with field viruses. According to Horiuchi⁷ and Stuetzer & Hartmann⁸ there have been changes in nonstructural protein 1 (NS1) and capsid 2 (VP2) genes based on time, then Garcia R, Castro TX & Miranda⁹ and Lin¹⁰ stated that antigenic variations of parvo viruses that attack dogs are identified by the names 2a, 2b, and 2c. Feline Panleukopenia Viruses have been found to infect several hosts with mutations in their capsid proteins.^{11,12} Protein capsid mutations in FPV result in changes in the amino acid cysteine into serine at position 273 which can cause infection in new hosts.^{13,14}

Table 1. Individual information on Panleukopenia Virus Samples in Cats

No	Name	Age	Gender	Origin	Symbol
1	<i>Felocell 3 Vaccine</i>	-	-	-	V1
2	<i>Purevax RCP Vaccine</i>	-	-	-	V2
3	Lilo	1 year	Female	East Java	V3
4	Happy	5 months	Male	East Java	V4
5	Miko	4 months	Male	East Java	V5
6	Samba	1 year	Male	East Java	V6
7	Molly	5 months	Female	East Java	V7
8	Bona	7 months	Male	East Java	V8

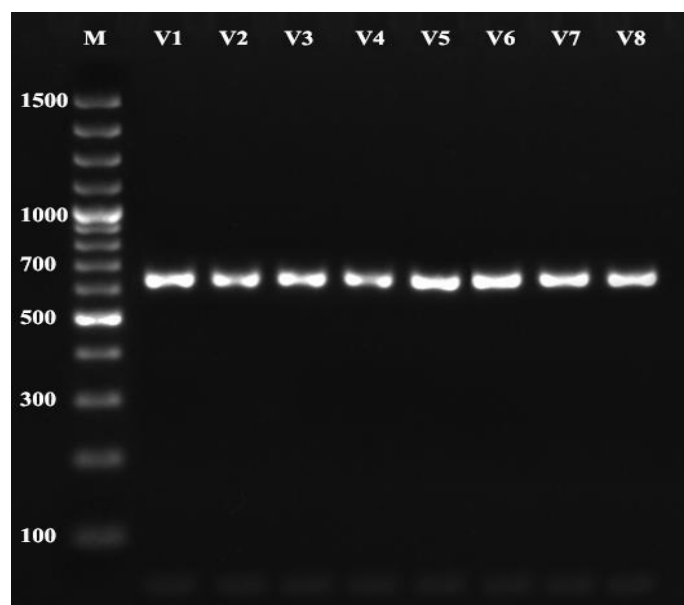


Figure 1: PCR results show a single band of ± 630 bp in the VP1 and VP2 genes according to the specifications of a pair of primers designed. M: 100bp Marker; V1: Felocell® 3 vaccine; V2: Purevac RCP vaccine; V3: Patient Lilo; V5: Happy Patient; V6: Samba patient; V7: Patient Molly; V8: Patient Bona

	10	20	30	40	50	60	70	80
FFV_Strain Philips Roxane	ATTCTCGCCAGCAGATCAACGCTTATAGATCAAACTAAGGACGCTAAGATTGGGGGGGAAATAGGACATTATTT							
FFV_Strain FFV-BJ05C.....							
FFV_Strain FFV-BJ04	...T.....A.....C.....							
FFV_Strain IZSSI 3201 1 15C.....							
FFV_Strain IZSSI 42807 15	...T.....							
FFV_Strain XJ-1							
FFV_Strain FFV/Raccoon/RC6/BC	--..T.....							
FFV_Strain FFV/Raccoon/RC18/BC	--..T.....							
FFV_V1 (Felocell 3)							
FFV_V2 (Purevax RCP)							
FFV_V3	...T.....							
FFV_V4	...T.....							
FFV_V5	...T.....							
FFV_V6	...T.....							
FFV_V7	...T.....							
FFV_V8	...T.....							
	90	100	110	120	130	140	150	160
FFV_Strain Philips Roxane	TTTAGAGCTAAAAAGCAATTGCTCCAGTATTAAGTATACACCCAGATCCTCCATCAACATCAAGACCCAGCAAAACCAAC							
FFV_Strain FFV-BJ05A.....							
FFV_Strain FFV-BJ04A.....							
FFV_Strain IZSSI 3201 1 15G.....A.....							
FFV_Strain IZSSI 42807 15G.....A.....							
FFV_Strain XJ-1G.....A.....							
FFV_Strain FFV/Raccoon/RC6/BCG.....A.....							
FFV_Strain FFV/Raccoon/RC18/BCG.....A.....							
FFV_V1 (Felocell 3)							
FFV_V2 (Purevax RCP)							
FFV_V3G.....A.....							
FFV_V4G.....A.....							
FFV_V5G.....A.....							
FFV_V6G.....A.....							
FFV_V7G.....A.....							
FFV_V8G.....A.....							
	170	180	190	200	210	220	230	240
FFV_Strain Philips Roxane	TAAAAGAGGTAAACCCACCTCTATTTTCATCAATCTTGCAAAAAAAAAAAGCCGGTGCAGGACAGTAAAAAGAG							
FFV_Strain FFV-BJ05							
FFV_Strain FFV-BJ04							
FFV_Strain IZSSI 3201 1 15							
FFV_Strain IZSSI 42807 15							
FFV_Strain XJ-1							
FFV_Strain FFV/Raccoon/RC6/BC							
FFV_Strain FFV/Raccoon/RC18/BCG.....							
FFV_V1 (Felocell 3)							
FFV_V2 (Purevax RCP)							
FFV_V3G.....							
FFV_V4G.....							
FFV_V5G.....							
FFV_V6G.....							
FFV_V7G.....							
FFV_V8G.....							
	250	260	270	280	290	300	310	320
FFV_Strain Philips Roxane	ACAATCTTGCAACCGATGAGTGATGGAGCAGTTCAACCCAGACGGTGGTCAACCTGCTGTGAGAAATGAAAGAGCTACAGGA							
FFV_Strain FFV-BJ05A.....							
FFV_Strain FFV-BJ04A.....							
FFV_Strain IZSSI 3201 1 15A.....							
FFV_Strain IZSSI 42807 15A.....							
FFV_Strain XJ-1A.....							
FFV_Strain FFV/Raccoon/RC6/BCA.....							
FFV_Strain FFV/Raccoon/RC18/BCA.....							
FFV_V1 (Felocell 3)A.....							
FFV_V2 (Purevax RCP)A.....							
FFV_V3A.....							
FFV_V4A.....							
FFV_V5A.....							
FFV_V6A.....							
FFV_V7A.....							
FFV_V8A.....							

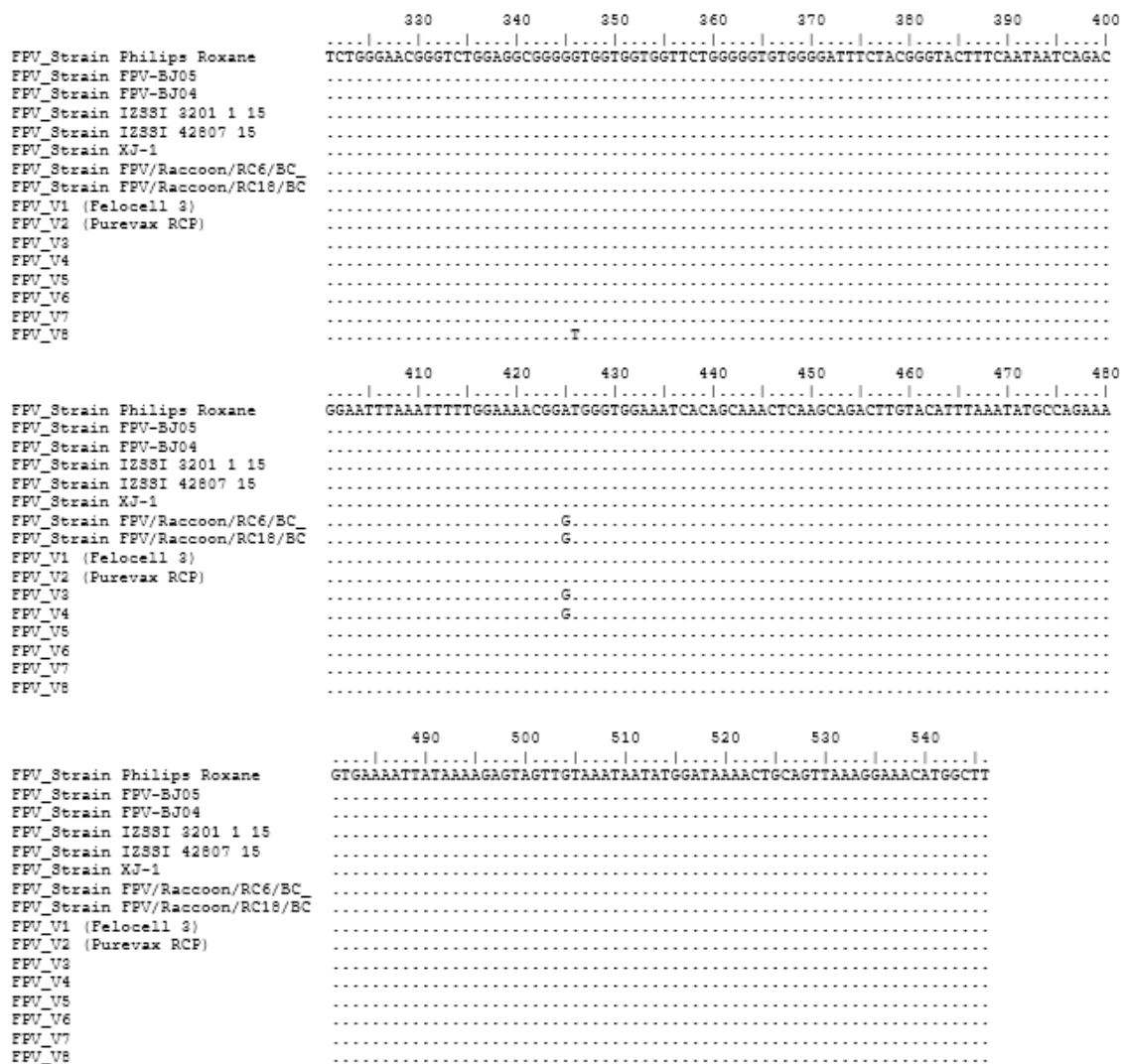


Figure 2: Alignment results of PCR product sequences of VP1, VP2 FPV genes to database of FPV strains according to NCBI

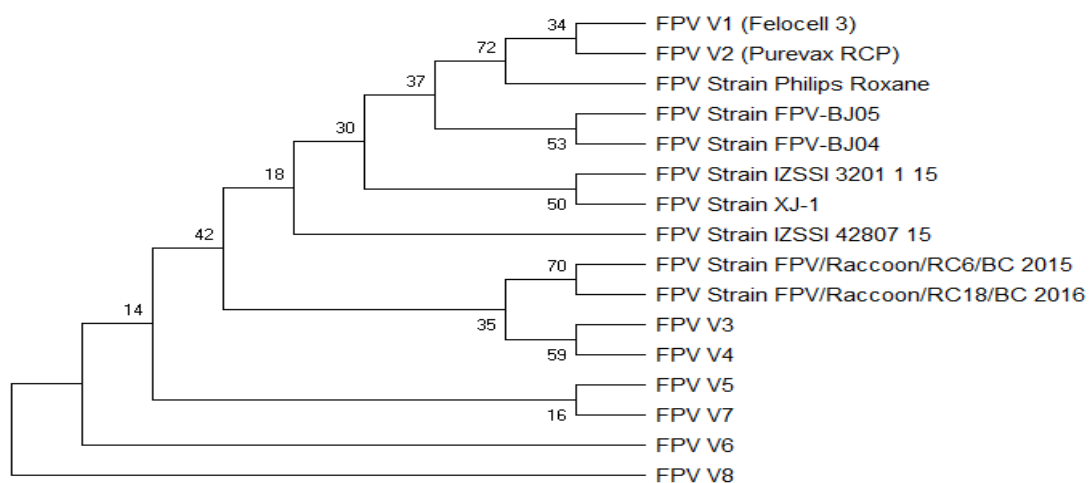


Figure 3: Genetic distances in viral DNA sequencing results based on virus strains in the database, vaccines and field isolates

FPV_Strain_Philips_Roxane															
FPV_Strain_FPV-BJ05	0.0055														
FPV_Strain_FPV-BJ04	0.0092	0.0037													
FPV_Strain_IZSSI_3201_1_15	0.0055	0.0037	0.0074												
FPV_Strain_IZSSI_42807_15	0.0074	0.0055	0.0055	0.0018											
FPV_Strain_XJ-1	0.0055	0.0037	0.0074	0.0000	0.0018										
FPV_Strain_FPV/Raccoon/RC6/BC_2015	0.0074	0.0055	0.0055	0.0055	0.0037	0.0055									
FPV_Strain_FPV/Raccoon/RC18/BC_2016	0.0112	0.0093	0.0093	0.0093	0.0074	0.0093	0.0037								
FPV_V1_(Felocell_3)	0.0018	0.0037	0.0074	0.0037	0.0055	0.0037	0.0055	0.0093							
FPV_V2_(Purevax_RCP)	0.0018	0.0037	0.0074	0.0037	0.0055	0.0037	0.0055	0.0093	0.0000						
FPV_V3	0.0111	0.0092	0.0092	0.0055	0.0037	0.0055	0.0037	0.0074	0.0092	0.0092					
FPV_V4	0.0111	0.0092	0.0092	0.0055	0.0037	0.0055	0.0037	0.0074	0.0092	0.0092	0.0000				
FPV_V5	0.0092	0.0074	0.0074	0.0037	0.0018	0.0037	0.0055	0.0093	0.0074	0.0074	0.0018	0.0018			
FPV_V6	0.0092	0.0074	0.0074	0.0037	0.0018	0.0037	0.0055	0.0093	0.0074	0.0074	0.0018	0.0018	0.0000		
FPV_V7	0.0092	0.0074	0.0074	0.0037	0.0018	0.0037	0.0055	0.0093	0.0074	0.0074	0.0018	0.0018	0.0000	0.0000	
FPV_V8	0.0111	0.0092	0.0092	0.0055	0.0037	0.0055	0.0074	0.0111	0.0092	0.0092	0.0037	0.0037	0.0018	0.0018	0.0018

Figure 4: Results of genetic distance between FPV strains with vaccines and field isolates using Pairwise Distance with Kimura-2 Parameter modeling

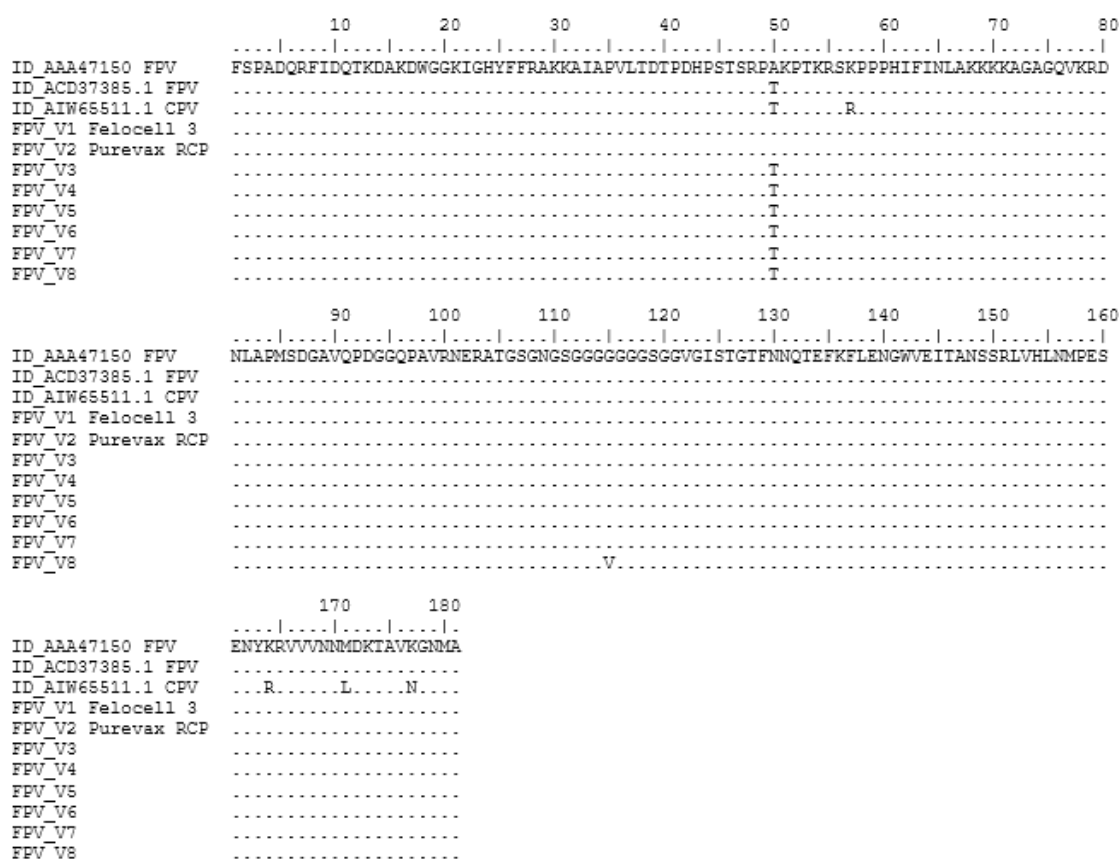


Figure 5. Analysis of Amino Acid in FPV and other isolates

Based on the alignment results, the results of the sample sequences of V1 and V2 were matched, respectively the Felocell® 3 vaccine and Purevax RCP. Whereas FPV virus field isolates namely V3 - V8 samples showed some differences in sequences namely in sequences 5, 95, 150, 173, and 425 (Figure 5). Differences in nucleotide bases in some strains, vaccines and field isolates also show a clear genetic difference in phylogenetic trees. Alignment of DNA sequences obtained homogeneous in several strains of FPV according to the NCBI database, which is precisely in the VP1 protein, VP2 FPV. The results of sequencing from eight samples showed a match on eight strains of FPV virus in a database with identities ranging from 99.2% - 99.8% (Figures 2.3 and 4).

The predominant vaccine against FPV is a modified live virus vaccine (MLV) which has been repeatedly discussed on feline and mink cell lines. Inactivated vaccines have been widely used in the past but provide low levels of protective antibodies, which have relatively short lives.^{15,16} Several studies have reported the occurrence of sero-negative kittens at the age of 15 weeks who have been vaccinated once or twice, and parvovirus disease in some vaccinated kittens shows that vaccination may not always be successful despite using standard protocols.^{17,18}

CONCLUSION

From the results of the research conducted, FPV isolates were obtained and could be used as vaccine candidates for FPV in Indonesia.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCE

1. Jakel V, Cussler K, Hanschmann KM, Truyen U, König M, Kamphuis E and Duchow K. 2012. Vaccination against Feline Panleukopenia: implications from a field study in kittens. *BMC Vet Res.*; 8: 62.
2. Truyen, U. And Parrish, C. R. 2013. *Feline Panleukopenia Virus: Its Interesting Evolution and Current Problems in Immunoprophylaxis Against a Serious Pathogen*. Vet. Microbiol, 165(1-2):29-32.
3. MacLachlan NJ, Dubovi EJ, Barthold SW, Swayne DF & Winton JR (2016) *Fenner's Veterinary Virology*, 5th ed., Academic Press, Elsevier, USA. ISBN 9780128011706.
4. Nakamura, K., Sakamoto, M., Ikeda, Y., Sato, E., Kawakami, K. & Miyazawa, T. et al., 2001, 'Pathogenic potential of canine parvovirus types 2a and 2c in domestic cats', *Clinical and Diagnostic Laboratory Immunology* 8(3), 663-668.
5. Ohshima T & Mochizuki M (2009) Evidence for recombination between feline panleukopenia virus and canine parvovirus type 2. *The Journal of Veterinary Medical Science*, 71: 403-408.
6. Kruse, B. D., S. Unterer, K. Horlacher, C. Sauter-Louis, and K. Hartmann. 2010. Prognostic Factors in Cats with Feline Panleukopenia. *J. Vet. Intern. Med.*; 24:1271-1276.
7. Horiuchi, M., Yamaguchi, Y., Gojobori, T., Mochizuki, M., Nagasawa, H., Toyoda, Y., Ishiguro, N. & Shinagawa, M. 1998. Differences in the evolutionary pattern of feline panleukopenia virus and canine parvovirus. *Virology* 249, 440-452.

8. Stuetzer B & Hartmann K (2014) Feline parvovirus infection and associated diseases. *Veterinary Journal*, 201: 150-155.
9. Garcia R, Castro TX & Miranda SC (2011) Characterization of parvoviruses from domestic cats in Brazil. *Journal of Veterinary Diagnostic Investigation*, 23: 951-955.
10. Lin, C.N., Chien, C.H., Chiou, M.T., Chueh, L.L., Hung, M.Y., Hsu, H.S. 2014. Genetic characterization of type 2a canine parvoviruses from Taiwan reveals the emergence of an Ile324 mutation in VP2. *Virology* 511:39.
11. Allison, A.B., Kohler, D.J., Ortega, A., Hoover, E.A., Grove, D.M. & Holmes, E.C. et al. 2014, 'Host-specific parvovirus evolution in nature is recapitulated by in vitro adaptation to different carnivore species', *PLoS Pathogens* 10(11), e1004475, <http://dx.doi.org/10.1371/journal.ppat.1004475>.
12. Sykes JE (2014) Feline Panleukopenia Virus Infection and Other Viral Enteritides. In: *Canine and Feline Infectious Diseases* ed. Sykes JE, Elsevier Inc., USA. ISBN 978-1- 4377-0795-3.
13. Greene CE (2012) Feline Enteric Viral Infections. In *Infectious Diseases of the Dog and Cat 4th ed.*, ed. Greene CE, Elsevier Inc., USA, ISBN 978-1-4160-6130-4.
14. Callaway HM, Feng KH & Lee DW (2016) Parvovirus capsid structures required for infection: mutations controlling receptor recognition and protease cleavages. *Journal of Virology*, 91: 1-16.
15. Gueguen S, Martin V, Bonnet L, Saunier D, Mahl P, Aubert A. 2000. Safety and efficacy of a recombinant FeLV vaccine combined with a live feline rhinotracheitis, calicivirus and panleukopenia vaccine. *Vet Rec*, 146 (13):380–381.
16. Gore TC, Lakshmanan N, Williams JR, Jirjis FF, Chester ST, Duncan KL, Coyne MJ, Lum MA, Sterner FJ. 2006. Three-year duration of immunity in cats following vaccination against feline rhinotracheitis virus, feline calicivirus, and feline panleukopenia virus. *Vet Ther*, 7(3):213–222.
17. Dawson S, Willoughby K, Gaskell RM, Wood G, Chalmers WS: A field trial to assess the effect of vaccination against feline herpesvirus, feline calicivirus and feline panleukopenia virus in 6-week-old kittens. *J Feline Med Surg* 2001, 3(1):17–22.
18. Zhang W, Li L, Deng X, Kapusinszky B, Pesavento PA & Delwart E (2014) Faecal virome of cats in an animal shelter. *Journal of General Virology*. 95: 2553-2564.