Detection of Metallo β-lactamase in *Klebseilla*pneumoniae Isolated From Upper Respiratory Tract of Carrier individuals In Iraq

Nadia Abdulrazzaq Jamal*1, Mohammed Nuh AL-Khafaf 2 and Saladdin M. Abdulazeez 3

1,2 Community Technique Department, Technical Institute of Mosul, Northern Technical University, Iraq

3 Medical Laboratory Techniques Department, Technical Institute Mosul, Northern Technical

University, Iraq

Corresponding Author: Nuhmohammed@ntu.edu.iq

ABSTRACT

Klebsiella pneumoniae is an a health care challenge over all the world, due to its resistance to many effective antibacterial, particularly those depend as a standard therapeutic options, Klebsiella carbapenemase producing have been emerged in Indian provinces, this phenomenon then have been disseminated in many countries $\ \$ around the world. The most successful $\ \beta$ -lactam hydrolyzing enzymes are belonging to the above antibiotic category is $\,$ New Delhi metallo $\beta\text{-lactamase},$ presence of NDM is an indication of multi-drug resistance. K.pneumoniae was isolated from Indian employees in Erbil city and from Iraqi people living in Mosul city, who had previously conducted an interventional surgery at hospitals in India.Of a total of 35 K.pneumoniae isolates, 6 KCP strains were discovered by phenotypic analysis using the modified Hudge test. While molecular screening showed 3 KCP as a true positive result. High molecular sensitivity and specificity, based on Gradiant Multiplex PCR, were found in the comparison with classical diagnosis method.On the other hand, according to the Random Amplification Polymorphic DNA, genetic variation has categorized the six KCP isolates into three clusters due to the existence and absence of random amplicons.

Keywords: Klebsiella pneumonia, NDM, Multiplex-PCR, RAPD-PCR.

Correspondence:

Nadia Abdulrazzag Jamal

Community Technique Department, Technical Institute of Mosul, Northern Technical University, Iraq

*Corresponding author: Nadia Abdulrazzaq Jamal email-address: Nuhmohammed@ntu.edu.iq

INTRODUCTION

Klebsiella pneumoniae resist large number antimicrobials and are widely recognized as a significant source of multidrug resistance genes that can disseminate among gram negative genera. Owing to its resistance to "ultimate option" antimicrobials such as last generation quinolones and carbapenems, it has gained notoriety and therapeutic choices are, in turn, limited here. ^(15,22,25). Carbapenem antibiotics belong to β-lactam antibiotics category, that are reliant on treating several infections caused by Multidrug resistant (MDR) bacteria. Enzymes are the most prevalent mechanism in the population of pathogenic bacteria involved in an emergence of carbapenem-resistant strains, due to inactivation of β-lactam molecules and occurrence of persistent resistance to β -lactam antibacterial strains. (3,13). β-lactamases are dividing into two sub groups serine βlactamase and Metallo β-lactamase(MBL). They were categorized into four groups according to the Ambler classification, three of them (A, C, D)classified as serine βlactamase as serine β-lactamase and B-class enzymes belonging to the Metallo β-lactamase Carbapenemase ezymes belonging to the classes A, B, and D⁽¹⁰⁾. New Delhi Metallo-β-Lactamase (NDM) is an Ambler class B, that firstly identified in K. pneumoniae and Escherichia coli during 2008 in Indian sub-continent. Later, NDM producers have been described worldwide. blaNDM-1 1 is an NDM-1 encoding gene nomenclature situated on a transmissible plasmid(20,26). NDM-1 was listed as the first source of new variants such as NDM-2 till to NDM-17, but all of them showed similar enzymatic activities against β-lactam antibacterial⁽¹⁰⁾.Antimicrobial susceptibility test and Huge test consider as perfect tool to observe carbapenem bacterial resistance, while the molecular screening is a confirmatory test to demonstrate responsible genes(1,4,6).

Several studies have been published on isolation and identification of carbapenemase producing Klebsiella (KCP) depending on phenotypic demonstration and molecular detection as a local documented research^(18,19,27). In addition to Hudson, Hu and Arpin as an International studies^(2,5,9,6).

The present study focuses on isolation of KCPs from certain individuals in Mosul society, who traveled to India for health care, in addition to Indian workers in Erbil city, as KCP carriers and transporters to our region. As well as to identification of the molecular relationship between isolated strains.

MATERIAL AND METHODS

Samples collection and bacterial identified

A total of 190 upper respiratory swabs (URSs) were obtained from healthy Indian workers(120),and surgical intervention patients(70)who were treated in India, from November 2017 till end of April 2018 Erbil and Mosul cities, respectively. Thirty five isolates of non-duplicated *K.pneumonia* were identified using standard bacteriological technique⁽⁷⁾.

Carbapenems Susceptibility Test

The Kirby-Bauer disc diffusion method was performed using Mueller Hinton agar medium supplied by Hi-media, India, to evaluate KCP susceptibilities to certain β -lactam antibiotics (imepenem and meropenem, resistance zone diameter, ≤ 21 mm) As outlined in detail by CLSI⁽⁶⁾.

Phenotypic Carbapenemase Production estimation

Carbapenemase production was evaluate by Hodge test using meropenem disc 10 μg . MBLs (blaNDM-1 and blaNDM-2 genes)production confirmed by synergistic reaction of 10 μg imipenem discs add to 290 μg EDTA (Sigma Chemicals, USA),which placed onto Mueller-Hinton agar plate after cultivation of bacterial strains by pour plate method. Inhibition zone increased around imipenem EDTA disc(diameter>4mm)in comparison with

imipenem disc alone, which was depended as visible indicator for production of $\,\mathrm{MBLs}^{(16,17)}.$

Molecular detection of β-lactamase genes:

Episomal DNA was extracted from previous positive screening isolates using spin plasmid basic, Canada) as manufacturer's protocol description, Amplification mixture was subjected to Gradiant thermocycler(Applied biosystem, USA) to demonstrated blaNDM-1 gene blaNDM-2 gene via multiplex PCR⁽⁸⁾.using the following specific primer GTCTGGCAGCA CACTTCCT-3')and sets, NDM-1-F(5'-NDM -1 -R(5'-CGTATGAGTGATTGCG GCG-3') with accession No.(KT956171) and NDM-5-F(ATCACGATCATGCTG GCCTT) and NDM-5-R(TTGCGACTTATGCCAATGCG)with accession No.(MK372393) documented by National Center for Biotechnology information, NCBI⁽²⁶⁾. Amplification program included 95 C as DNA unwinding for 180 sec and 35 times of each of denaturation 94 C for 30 sec, hybridization 55 and 54 C(respectively) for 30 sec, extension 72 C for 30 sec, in addition to final extension72 C for 180sec, for both two nrimer sets

Genetic diversity of KCP

Extraction of genomic DNA was conducted as that detailed by promega company, gDNA concentration and purity were carried out by nano-spectrophotometer, Template integrity was estimated according to that documented by⁽²⁴⁾.KCP variety was identified via Randomly amplification polymorphic DNA(RAPD) for amplification of four random primers (Bioneer, S.Korea), which were included OPAX-01(GTGTG CCGTT),OPAV-08 (TGAGA AGCGG),OPBH-17(CTCTTACGGG) and OPAT-01(CAGTGGTTC C) . Primex was supplied by Bioneer

company, amplification mixture was placed in Gradiant thermocycler (Applied biosystem, USA)wells to carry out the following program: Pre-incubation 94 C for 240s,followed with 35 cycles of 93 C for 30s as initial incubation,33,31,27 and 28 C respectively for 30s as primers hybridization temperature and 72 C for 30s as complementary DNA strand extension, then finally 72 C for 180s as a terminal extension.

Outcomes documentation of Genotyping and Genetic diversity analysis

All amplification products were detected and photographed using gel documentary (ATTA,Japan), all DNA bands were classified to polymorphic and single, they were represented as twodigital data(1,refere to band;0,absence) Then, we calculated a simple matching (Sm) coefficient and a Jaccard algorithm to determine a dendrogram and similarity and dissimilarity. (PASS Software, Oslo,2009). The specificity, sensitivity and diagnostic accuracy of the results were also calculated by applying the following equations: $(a / a + b) \times 100 = \text{sensitivity}, (d / d + c) \times 100$ = specificity⁽²³⁾.

RESULTS

Bacterial isolation

The present results showed that the percent of *K. pneumoniae* isolation from URSs was 18.4%,35/190(Table1). This finding was nearly to that reported by Bora and Ahmed(2012), who showed that 19/219(8.7%) isolates were revealed positive results for NDM-1 production.

Table (1): Number and percentage of K. pneumoniae isolation from URSs.

Isolation sources N		Positive results No. (%)	Negative results No. (%)			
Clinical samples	190	35(18.4)	155(81.6)			
Cal. $X^2 = 0.82$, tab. $X^2 = 0.604$, df = 1, significance P<0.01.						

Biochemical reaction revealed that all K. pneumoniae isolate showed mucoid lactose fermentation colonies (pink color)on Macconkey agar, with oxidase negative reaction and Gram's negative rods under oil immersion, while IMViC (Indol, Methyl red, vogas proskuaer and Citrate tests) revealed negative result for I and M and positive result for Vi and $M^{(7,21)}$.

Phenotypic and genotypic screening of the Carbapenemase Production

Of all 35 *K. pneumoniae*, 6(17.1%) isolates showed positive appearance towards Hodge test, but PCR amplification products of *bla*NDM-1 gene and *bla*NDM-2 gene revealed 3(8.6%) isolates with an amplicon size 640 base pair(bp.) and 314bp. respectively associated with Indian workers samples(Figure 1, table 2).

Gene	Indian workers	Tract of C Iraqian Citizen	Tests	Sensitivity(%)	Specificity(%)
blaNDM-1,2 genes	3		PCR method	50	50
	4	2	Hodge test	100	100

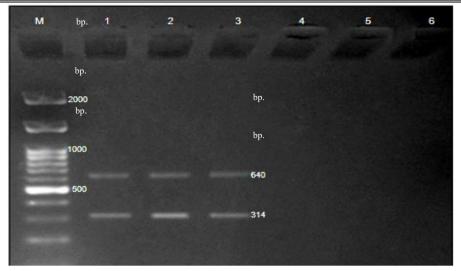


Figure (1): Multiplex amplification products for *bla*NDM-1 and *bla*NDM-2 genes from *K. pneumoniae* isolates. Lanes M, marker 100-bp, lanes (1-3) Amplicon size 640 bp. for *NDM-1* and 314 bp. for *NDM-2*, lanes (4-6) showed negative results.

Our results were similar to that finding by⁽²⁰⁾.who found that 15/275(5.5%)isolates of *K. pneumoniae* revealed

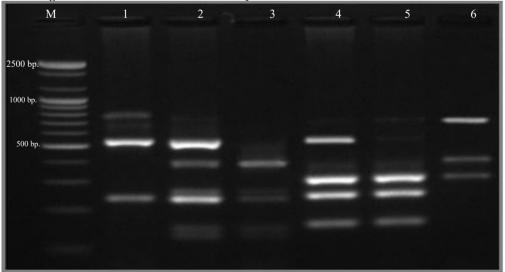
positive outcome for NDM-1 gene Table(2). Other study carried out by $\operatorname{Hasan}^{(18)}$, showed that 14(10.9%) isolates of K. pneumonia were carbapenemase producers. On the other hand, phenotypic investigation and molecular screening showed high specificity and sensitivity of PCR in comparison with modified Hodge test results. Conventional PCR is a sensitive and accurate technique, which need significant requirements to be optimized⁽¹⁰⁾.

Table 2: comparison between modified Hodge test and PCR results

Genetic diversity of KCP

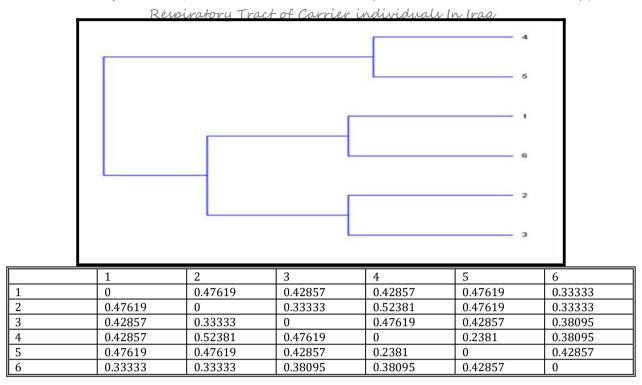
Figure two represented gDNA fragments for six KCP isolated strains, genetic diversity revealed high similarity with low distance indices between the strains 4 and 5, whereas high distance indices and low similarity

were noticed between 2 and 4 strains(Table 2).Random amplification polymorphic DNA is an essential tool to determine the genetic relationship among microorganisms⁽¹⁴⁾.



Figure(2):Random PCR amplicons for six KCP isolates. Line M is a DNA marker (100bp.), Lines 1-6 are a random gDNA fragments for KCPs

Table (3): KCP similarity and dissimilarity



The dendrogram among the phenotypically diagnosed KCPs showed genetic relationship with two mainly clusters, the first is single have two isolated strains(4 and 5), the second have two sub divisions each of them

represented with two isolated strains 1,6 and 2,3 respectively(Figure 3).Random amplification of gDNA is an essential tool to detect the genetic diversity among microorganisms⁽¹²⁾.

Figure (3): Phylogenetic tree of six KCPs

RECOMMENDATION

Depending on our result we recommended that

- Iraqi government must depend clinical testing program for checking all incomes passengers to the country at the borders, in order to control the administration and prevention of non endemic diseases.
- 2. Recommended for all hospital especially intensive care units to check out medical staff for detect any variant strains they which carrier them, in order to maintain hospital care units clear from any health care problem.

REFERENCES

- Amjad A, Mirza I A, Abbasi S A, Farwa U, Malik N, and Zia F. Modified Hodge test: a simple and effective test for detection of carbapenemase production. Iran J Microbiol. (2011);3(4):189-193.
- Bora A and Ahmed G. Detection of NDM-1 in Clinical Isolates of Klebsiella Pneumoniae from Northeast India. Journal of Clinical and Diagnostic Research. (2012);6(5):794-800.
- 3. Bush K, Jacoby GA. Updated functional classification of beta-lactamases. Antimicrob Agents Chemother. (2010);54:969–76.

- Carvalhaes C, Picao R, Nicoletti A, Xavier D. and Gales A. Cloverleaf test(modified Hodge test) for detecting carbapenemase production in *Klebsiella* pneumoniae:be aware off alse positive results. J. Antimicrob. Chemother.(2009);65:249-251.
- Celikbilek N,Unaldi O, Kirca F, Gozalan A and Acikgoz Z. Molecular characterization of carbapenemresistant Klebsiella pneumoniae species isolated from a Tertiary Hospital, Ankara, Turkey. Jundishapur J. Microbiol. (2017):10(10):120-123.
- Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Ninth Informational Supplement. (2019); CLSI document M100- S29. Wayne, PA: Clinical and Laboratory Standards Institute.
- Collee J G, Miles R S, Wan B .Tests for the identification of bacteria. *In*: Collee J G, Fraser A G, Marmion B P .Mackie and McCartney Practical Medical Microbiology, 14th ed. Edinburgh: Churchill Livingstone. (1996); 131-50
- 8. Dallenne C, Dacosta A, Decré D, Favier C and Arlet G. Development of a set of multiplex PCR assays for the detection of genes encoding important betalactamases in Enterobacteriaceae. J. Antimicrob. Chemother. (2010); 65:490-495.

- Dash N, Panigrahi D, Zarouni M A, Darwish D, Ghazawi A, Sonnevend A, Pal T, Yasin F and Hadi S A. High incidence of New Delhi metallo-β-lactamase producing *Klebsiella pneumoniae* isolates in Sharjah, United Arab Emirates. Microbial Drug Resistance.(2014);20:52-56.
- 10. Dhillon R H and Clark J .ESBLs: A clear and present danger? Critical Care Research and Practice.(2018);2090-1305.
- Dillon B, Thomas L, Mohmand G, Zelynsky A and Iredell J. Multiplex PCR for screening of integrons in bacterial lysates. J. Microbiol. Methods.(2005);62:221-232.
- Drancourt, M.; Bollet, C.; Carta, A. and Rousselier, P. Phylogenetic analyses of Klebsiella species delineate Klebsiella and Raoultella gen. nov., with 201 description of R. ornithinolytica comb. nov., R. terrigena comb. nov. and R. planticola comb. nov. Intern. J. Systematic and Evolutionary Microbiology.(2001);51: 925-932.
- 13. El-Gamal M I, Brahim I, Hisham N, Aladdin R, Mohammed H and Bahaaeldin A. Recent updates of carbapenem antibiotics. European J. Medicinal Chemistry. (2016);31:185-195.
- Elina B Reinoso and Susana G Bettera. Random Amplified Polymorphic DNA PCR in the Teaching of Molecular Epidemiology. Biochemistry and Molecular Biology Education. (2016);44(4) Pages 391–396.
- Firoozeh F, Corehtash Z G, Khorshidi A, Akbari H and Aznaveh A M. Biofilm formation and virulence factors among *Klebsiella pneumoniae* isolated from burn patients. Jundishapur J. Microbiol.(2019); 8(10):118-122.
- Franklin C, Liolios L, Peleg AY. Phenotypic detection of carbapenem susceptible metallo-beta-lactamaseproducing gram-negative bacilli in the clinical laboratory. J Clin Microbiol. (2006); 44(9): 3139 -44
- 17. Girlich D.; Poirel, L. and Nordmann, P. Value of the modified Hodge test for detection of emerging carbapenemases in Enterobacteriaceae. J. Clin. Microbiology, (2012). 50: 477-479.
- 18. Hassan H, N A Yassin, A T Saadi. Molecular Detection of the Genes bla OXA, bla KPC and bla NDM Among Carbapenem Resistant *Klebsiella Pneumoniae* Isolated From Different Hospitals in Duhok City (2019); The Internet Journal of Microbiology.17.1.
- Jaber H M R. Phenotypic and genotypic detection of antibiotic resistance Klebsiella pneumoniae in AL-Najaf Governorate .Iraq. (2012), MSc. Thesis. College of Science. Kufa University.
- 20. Kanj S S and Kanafan Z A. Current concepts in the antimicrobial therapy against resistant gramnegative organisms: extended-spectrum beta lactamase producing Enterobacteriaceae, carbapenem resistant Enterobacteriaceae and multidrug resistant *Ps. aeruginosa*. Mayo. Clin. Proc. 2011; 86(3):250-59.
- MacFaddin J F. Biochemical tests for identification of medical bacteria. 3rd. ed. Lippincott Williams and Wilkins, USA. Microbiology. 2019;17.1.
- 22. . Munoz Price L S, Poirel L, Bonomo R A, Schwaber M J, Daikos G L, Cormican M, Cornaglia G, Garau J, Gniadkowski M, Hayden M K, Kumarasamy K, Livermore D M, Maya J J, Nordmann P ,Patel J B, Paterson D L, Johann P, Villegas M V, Wang H,

- Woodford N and Quinn J P..Clinical epidemiology of the global expansion of *Klebsiella pneumoniae* carbapenemases. Lancet Infection Dis.(2013);13: 785-796.
- Niazi A D. Statistical Analysis in Medical Research. Republic of Iraq. Al-Nehrein University. (2000);148.
- Sambrook, J. and Russell, D. Molecular Cloning: A Laboratory Manual. (2001); 3rd. ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY,USA.
- 25. United States Department of Health and human Services Centers for Disease Control and Prevention. (2013). Antibiotic resistance threats in the United States.
- 26. Yam W K, Wailan A and Alikhan N F, *et al.* Complete plasmid sequence of *blaNDM-1* carrying IncA/C plasmid from Australia. (2013);NCBI.
- Zeinah R, Hameed Al-Sultani, Hadi R Rasheed Al-Taai. Detection of *NDM-1* in Cabapenem-Resistant *Klebsiella pneumoniae*. J. Pharm. Sci. & Res. (2019); 11(3):869-878.