Effect of Application of BMP2/TGF β1 in Traumatic Pulp of Osteoporotic Rat

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ABSTRACT

Introduction: Dental pulp tissue contains many undifferentiated mesenchymal cells, which have the ability to differentiate into different specialized cells. Induced pluripotent stem cells have been developed by various growth factors. The present study was designed to evaluate the effect of application of a combination BMP2/TGF β1 as capped material for traumatic pulp in osteoporotic rat.

Materials and Methods: Twelfth female rats (6 normal rat and other 6 osteoporotic rat), their maxillary anterior teeth subjected to mechanical traumatized pulpotomy, the left tooth has spared without treatment, while the right tooth capped with application of 0.5 μl of BMP2 and 0.5μl of TGF β1. Evaluation of histological changes includes scoring of pulp inflammation and scoring of morphology and thickness of dentin bridge were estimated for all study groups.

Results: Histological examination of tooth with pulptomy capped by BMP-2&TGF β1 for both normal and osteoporotic rat showed formation of reparative dentin bridge and minimal inflammatory response with a significant differences value in comparison to control.

Conclusion: The study concludes that application of a combination of BMP-2&TGF β1 enhanced tooth repair in osteoporotic rat.

Keywords: BMP-2, osteoporosis, pulpotomy, reparative dentin, TGF β1, tooth repair

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INTRODUCTION

Growth factors and extracellular matrix molecules, that are expressed during tooth development, are re-expressed in dental tissues under pathological conditions such as caries and dental injuries. They are often act as bio-stimulus to the pulp cells which can be differentiated to odontoblast1,2 or enhance progenitor cells to differentiate into odontoblast-like cells and produce a reparative dentin.3,4 Direct pulp capping by bioactive molecules or seeding of these molecules in the pulp may stimulate the formation of reparative dentin and enhanced pulp mineralization. Pulp-capping studies with different biological agents, including growth factors and extracellular matrix molecules, showed the formation of tertiary dentin (reparative dentin formation).5,6 Osteoporosis, is a prevalent metabolic bone disorder with increased susceptibility to fractures. It is also, thought to delay or impair the regenerative response.7,8 Osteoporotic fractures remain challenging to treat. Among other risk factors, decreased expression of morphogenetic proteins has been identified in osteoporosis.9,10 Studies on osteopontic animals show atrophy of the periodontal ligament (PDL) and that this atrophy was accompanied by a reduction in the pool of osteo-progenitor cells that associated with significantly slower extrusion socket healing.11,12 Bone morphogenetic proteins BMPs are group of bone-inducing growth factors that used to enhance osseous repair. Many studies have indicated that BMP promotes a proliferation and differentiation of dental pulp cells both in vitro and in vivo. Furthermore, BMP is used as a pulp capping agent, for its ability to induce the formation of reparative dentin.13,14 It was concluded that TGF-β1 could induce odontoblast differentiation and dentin formation and act as substrate for the progenitor cells to anchor and initiate the differentiation to odontoblast cells.15,16 The present study was designed to find the effect of using a combination of BMP-2 and TGF β1 as a new pulp capping material to enhance tooth healing in osteoporotic rats.

MATERIALS AND METHODS

Twelfth 8-week-old female wistar albino rats, weighed 230-250 g have been enrolled in this study and according to ethical approval from the ethical committee of the Al-Mustaqlab University College (license No: 069220) authorized all of the experimental approaches. These animals were kept in the animal department of National Center of Drug Control and Research (Iraq) at a constant humidity and temperature of 23°C and according to the National Council’s guide for the care of laboratory animals. A combination of Bone morphogenetic protein-2 and Transforming growth factor beta-1 factor were used as pulp capping material for maxillary anterior teeth of the normal and osteoporotic rats following the mechanical traumatized procedure. The control and experimental teeth have studied histologically (H&E stain) after 28 days postoperatively. Six teeth were enrolled from each of following groups.

Group 1: Normal rats with the pulptomy restored with resins glass ionomer cement.
Group 2: Normal rats with pulptomy capped by application of BMP-2&TGF β1.
Group 3: Induced osteoporotic rats with pulptomy restored with resins glass ionomer cement.
Group 4: Induced osteoporotic rats with pulptomy capped by application of BMP-2&TGF β1.

Induction of osteoporosis
Six rats were induced for osteoporosis by bilateral ovariectomy, and after 2 weeks postoperatively, the animals were received a systemic daily 1M injection of methylprednisolone hemi succinate (MPH) at dose (1 mg/kg) for 4 consecutive.
Operative procedure
After six weeks of ovariectomy, operative preparation was started. Before the pulptomy procedure, the experimental teeth were submitted to prophylaxis, and absolute isolation of the operative field was performed with a rubber dam. The tooth was washed with 0.2% chlorhexidine prior to cavity preparation. Then a classic class V cavity was prepared at the buccal aspect of the crown near the gingiva by using a sterile 0.8 mm slow-speed round diamond bur. The depth of the cavity was approximately half the diameter of the bur. Each bur was changed after 2 cavity preparations. During the preparation, the cutting area was irrigated with copious saline solution (0.9% NaCl) to prevent heat generation. Then pulp exposure were induced using dental explorer in the middle point of the cavity (Shayegan et al, 2012).17 Pulpal bleeding was controlled by pressing with a sterile saline-soaked cotton pellet and paper points. The left tooth was considered a control, represented the group 1&3 and only a cotton pellet was inserted over the exposed pulp tissue and the cavity was restored with resinous glass ionomer cement (Vitremer, 3M ESPE, St. Paul, Minn., USA). The right tooth was considered an experimental, represented the group 2&4 with application of 0.5 μl of BMP2 (rhBMP-2) Medtronic Sofamor Danek;TN/USA and 0.5μl of TGFβ1 (ab50036,Abcam) was placed in the pulp chamber using micro-pipette, allowed for one minute then restored, as previously described.

Histological Examination
At 28 days after direct pulp capping, the animals were sacrificed by an overdose of carbon dioxide gas. The whole maxilla was collected and fixed in 10% neutral buffered formalin for 24 hours at room temperature, and demineralized in 20% formic acid for 3 days. Then, the maxilla was sectioned carrying two anterior teeth, washed in 0.8 mm slow-speed saline solution (0.9% NaCl) to prevent heat generation. Then pulp exposure were induced using dental explorer in the middle point of the cavity (Shayegan et al, 2012).17 Pulpal bleeding was controlled by pressing with a sterile saline-soaked cotton pellet and paper points. The left tooth was considered a control, represented the group 1&3 and only a cotton pellet was inserted over the exposed pulp tissue and the cavity was restored with resinous glass ionomer cement (Vitremer, 3M ESPE, St. Paul, Minn., USA). The right tooth was considered an experimental, represented the group 2&4 with application of 0.5 μl of BMP2 (rhBMP-2) Medtronic Sofamor Danek;TN/USA and 0.5μl of TGFβ1 (ab50036,Abcam) was placed in the pulp chamber using micro-pipette, allowed for one minute then restored, as previously described.

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The stained sections were blindly evaluated by a trained investigator who was previously calibrated with an experienced pathologist. Under a light microscope (Olympus BX53, Olympus, Tokyo, Japan), all samples were evaluated and scored in terms of: 1) inflammatory cell infiltration, 2) reparative dentin formation, 3) reparative dentin bridge formation, and 4) odontoclast activity.

According to Mestrener et al (2003)14, the quantitation of intensity of inflammatory response and the thickness of dentine bridge were evaluated by counting them in visual field (X10, X20, X40) with subsequent of arithmetic mean for each specimen as follow:

<table>
<thead>
<tr>
<th>Intensity of inflammatory reaction</th>
<th>1. Absent or very few inflammatory cell.</th>
<th>II. Mild average number less than 10 inflammatory cells.</th>
<th>III. Moderate average number 10-25 inflammatory cells.</th>
<th>IV. Severe average number greater than 25 or necrosis.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thickness of Dentine Bridge</td>
<td>I. Up to 250μm.</td>
<td>II. 150- 249μm.</td>
<td>III. 1-149μm.</td>
<td>IV. Partial or absent bridge</td>
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STATISTICAL ANALYSIS
All records were entered into Excel spread sheets for evaluation with PASW statistics for windows, version 18.0. Statistical evaluation was executed using the SPSS (Statistical package deal for Social studies, Chicago, IL, united states of America). Descriptive statistics
1. Means
2. Frequency
3. Percentage

Inferential statistics
1. Chi square: to compare the scores of inflammatory response and thickness of dentin bridge between groups of teeth.
2. Likelihood ratio test: used as alternative to Chi square when the expected frequency is less than 1 in any cell or less than 5 in 20% of the cells.

RESULTS
Results for histological examination revealed that dentine-pulp interface for normal rats with pulptomy at 28th day showed thin layer of reparative dentin with inflammatory cells in sub odontoblastic area, while the view was differed for pulptomy capped with BM-P-2& TGFβ1, the reparative dentin was regular and mostly score II in thickness and dentin bridge formation was observed. Pulp shows odontoblast like cells occupy it's surface as flat cell with odontoblastic process extend in dentin. Calcospherite is detected as mineralized globules in reparative dentin, figure 1.

Histological examination for osteoporotic rats with pulptomy at 28th day showed internal dentin resorption with identification of odontoclast cells as multinucleated giant cell occupied the Howship's lacunae, the pulp shows fibrosis with fibroblast subside in dental pulp tissue and infiltration of inflammatory cells. Furthermore, Pulp shows a necrosis with no evidence of reparative dentin formation. Histological features for osteoporotic rats with pulptomy capped with BM-P-2&TGFβ1 at 28th day showed a formation of irregular reparative dentin with dentin bridge while pulp reveal an active new blood vessels with presences of inflammatory mononuclear cell infiltrated pulp tissue, figure 2.

Statistical results
Statistical analysis revealed a significant difference in frequency and percentage of inflammatory response, and the scoring of reparative dentin bridge thickness (μm) for the normal and osteoporotic with pulptomy capped by a combination of BMP2&TGFβ1 in comparison to control, tables (1,2).
Figure 1: Histological description for dentin-pulp interface for normal rats with pulptomy (untreated and treated) at 28th day

1A: Dentin-pulp interface for normal rats with pulptomy shows thin layer of reparative dentin (RD), inflammatory cells in subodontoblastic area (arrow). H&E x10

1B: Formation of dentin bridge (DB) for normal rats with pulptomy treated with BMP-2&TGFβ1. H&E x4

1C: View for shows reparative dentin (RD), odontoblast like cell (yellow arrows), odontoblast process (red arrows), calcosphere (white arrows) in normal rat with treated pulptomy. H&E x40

Figure 2: Histological description for dentin-pulp interface for osteoporotic rats with pulptomy (untreated and treated) at 28th day

2A: Dentin-pulp interface for osteoporotic rats shows internal resorption with odontoclast (arrows), pulp illustrates fibrosis (FB). H&E x20; 2B: Magnifying view for the pulp shows fibrosis (FB), necrosis (N), fibroblast (red arrows), inflammatory cells (black arrows). H&E x40; 2C: Dentin-pulp interface for osteoporotic rats with pulptomy treated with BMP-2&TGFβ1 shows dentin (D), reparative dentin (RD), demarcated line (red arrows), pulp (P) with inflammatory cells (black arrows). H&E x10; 2D: Magnifying view for the pulp shows active new blood vessels formation (BV), mononuclear cells infiltration (arrows). H&E x20; 2E: Formation of dentine bridge (DB). H&E x10

Table 1: Frequency and percentage of inflammatory response in different groups
DISCUSSION
The present study shows that application of a combination of BMP-2 and TGF-β1 as capped material for pulptomy induce new differentiation of mesenchymal cells into odontoblast like cell that apposed reparative bridge dentin with process of mineralization that appeared a calcospherite in both normal and osteoporotic rats. Mineralization of dental pulp was increased by TGF-β induction and increased the ALP activity of dental pulp cells as well as expression of dentin sialoprotein (DSP), osteopotin (OPN) and type I collagen. Our results coincide with different studies that shows the effect of using TGF-β and then apposition of reparative dentin. Many in vitro and in vivo studies illustrated the inductive potential of scaffold material combing with TGF-β1 to induce odontoblast differentiation and dentin formation from dental pulp cells. Other studies found that BMP-2 provides a strong signal for differentiation and mineralization of odontoblasts and osteoblasts and an over expression of BMP2 can promote fracture healing and osteogenic ability in senile osteoporotic fractures through activating the BMP/Smad signaling pathway. Animal studies related to post-menopausal estrogen deficient osteoporosis had shown healing to be prolonged with decreased levels of mesenchymal stem cells (MSCs) and decreased levels of angiogenesis, while in our study the application of combination of BMP-2 & TGF-β1 illustrates that many events has been affected including the recruitment and differentiation of (MSCs) to odontoblast like cell during the early phase; and angiogenesis with new blood vessels formation during pulp healing ; and finally formation of a reparative dentin.

CONCLUSIONS
The present findings implied that application of combination of BMP-2 & TGF-β1 in direct pulp capping for osteoporotic pulp healing, act as inductive agent that initiates the differentiation of odontoblast like cell and formation of dentin bridge and could be taken into consideration when designing capping material for inducing dentin tissue engineering. Further researchers should focus on this important topic and provide more data in this field in order to enable a sound clinical use of these materials in osteoporotic subjects.

ETHICAL CLEARANCE
all work of this study had done according to the National Council’s guide for the care of laboratory animals.

CONFLICT OF INTEREST
The authors declare that they have no conflict of interest.
REFERENCES


