

# Imino-Aldehydes-A New Tool to Combat Cancer and Viral Diseases

Uriel Bachrach\*

Department of Molecular Biology, Hebrew University-Hadassah Medical School, Jerusalem, Israel

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## ABSTRACT

The naturally occurring polyamine-spermine can be oxidised by serum amine oxidase to yield imino-dialdehydes. Similar dialdehydes can be synthesized chemically by treating diamines with chloro-propion-aldehydes. The oxidation products are cytotoxic and inhibit the growth of cancer cells as well as human, animal and plant viruses. A synthetic imino-dialdehyde, containing a diamine-hexane moiety, was more active than the spermine oxidation product and inhibited the growth of influenza and Newcastle disease viruses. The positively charged imino-dialdehydes, cross cellular membranes, interact with cellular nucleic acids and form a biologically inactive complex. Viral vaccines are usually prepared by treating viruses with formalin, which interact with viral membrane proteins. Human and or animal viruses treated with

imino-dialdehydes, retain viral membranes and are therefore potent immunogens. Vaccination of animals with influenza or Newcastle disease viruses, inactivated by imino-dialdehydes, gave better results compared to conventionally prepared vaccines. Cancer cells are rich in polyamines and treating these cells with amine-oxidases, results in the formation of cytotoxic imino-aldehydes. This approach can open new avenues in cancer chemotherapy, inactivating viruses and potent vaccines.

**Keywords:** Polyamines, Ornithine decarboxylase, Cancer, Imino-aldehydes

\***Correspondence:** Uriel Bachrach, Department of Molecular Biology, Hebrew University-Hadassah Medical School, Jerusalem, Israel, E-mail: urielb@ekmd.huji.ac.il

## INTRODUCTION

Ever since the discovery in the eighteen century, spermidine and spermine and their diamine precursor, putrescine, have been associated with cell growth. They are aliphatic amines which are positively charged and are known collectively as the polyamines. The link between polyamines and cancer was well established when polyamines were detected in Ehrlich ascites cells (Bachrach U, *et al.*, 1967) and elevated urinary polyamines were found in patients with a variety of tumors (Russell DH, 1971). As critical factors for cell growth and development, it is not surprising that these compounds are detected in tumor cells.

Ornithine Decarboxylase (ODC), the first enzyme in the polyamine biosynthetic pathway, exhibits increased enzyme activity in several cancers including breast prostate and colon (Russell DH and Snyder SH, 1968). Given the strong association between increased polyamine concentrations, elevated ODC activity and cancer, it was of interest to see if ODC inhibition could block cancer development.

## MATERIAL AND METHODS

### Synthesis of macromolecules

*Escherichia coli* was grown in glycerol-lactate medium and cells were incubated with H<sup>3</sup>-labelled uridine, thymidine or lysine. Reaction was stopped by the addition of trichloroacetic acid and the radioactivity in the precipitate was determined.

### Detection of ODC

The presence of ODC in individual cells was quantitatively detected by an immune-fluorescence assay, using an ACAS 570 computerized fluorescence microscope. The enzyme was also detected by a quantitative immune-histochemical assay, using ODC antibody and a Fluorescein Isothiocyanate (FITC)-linked antibody.

### Synthesis of imino-aldehydes

Imino-aldehydes are produced by incubating demines with chlor-

al-propionaldehyde-diethyl acetal of the oxalic acid salt. The diacetal was converted into the free aldehyde by incubating for 3 hours at 37°C with 0.05 N Sulphuric acid (H<sub>2</sub>SO<sub>4</sub>), followed by neutralization with 1 N Sodium Hydroxide (NaOH).

### Solubilized Sendai viruses

Sendai virus particles were solubilized by detergents like Triton X-100. The nuclear fractions in the solubilized mixtures are removed by centrifugation and amine oxidases are added to the fractionized membranes. Dialysis, removed the detergents and stable particles containing amine oxidases, were obtained.

## RESULTS AND DISCUSSION

### Targeting the polyamine pathway and chemotherapy

$\alpha$ -Difluoromethyl Ornithine (DFMO) was one of the first inhibitors (Figure 1) of ODC activity (Takigawa M, *et al.*, 1983). Despite its success *in vitro* in several tumor cell types, it proved to be less effective *in vivo* and in clinical trials. Apparently, compensatory mechanisms occur to increase polyamine cellular pools. On the other hand, DFMO can be used to prevent diseases (chemoprevention).

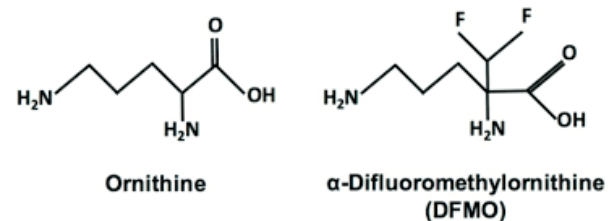


Figure 1: ODC and its inhibitor

### Amine oxidases

Polyamines are oxidized by various enzymes derived from different sources, including plants, pea seedlings and bacteria. Of all these enzymes, plasma amine oxidase has been the subject for the most extensive studies. The properties of this enzyme, its purifica-

tion and the possible functions of amine oxidases were discussed in great details by Tabor CW, *et al.*, 1954.

The plasma or serum of many ruminants, contain an enzyme which oxidized polyamines and benzylamine (but not diamines). The mechanism of polyamine oxidase by plasma amine oxidase was studied by Tabor CW, *et al.*, 1964 (Figure 2). Because of the liability of the oxidation products (Figure 2), they have to be used immediately after production or be stabilized. To study the biological activity of the oxidation product, catalase should be added to the system, to eliminate the presence hydrogen peroxide.

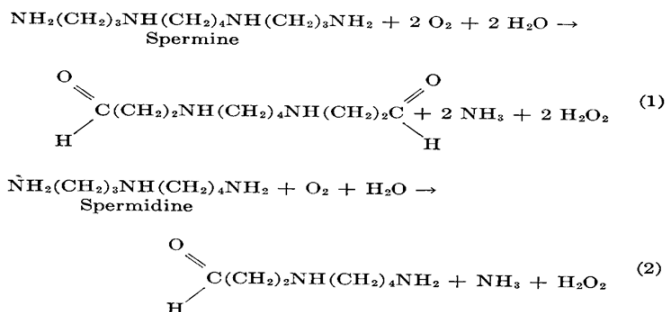


Figure 2: Oxidation of polyamines by serum amine oxidase

**Imino-aldehydes**

POX-3, (Figure 3), in the form of diacetal of oxalic acid salt was synthesized in our laboratory by Dambrovitz A. or purchased from Fine Organics Inc. Lodi, New Jersey, The diacetal was converted into the free aldehyde by incubating at 37°C for 3 hours with 0.05 N H<sub>2</sub>SO<sub>4</sub>, followed by neutralization with 1 N NaOH.

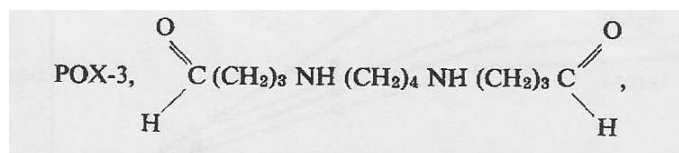


Figure 3: Diacetal oxalic acid salt

**Bacteria**

*Escherichia coli* was grown to a density of 2 × 10<sup>8</sup> cells/ml, incubated for 25 minutes with the imino-aldehyde (POX-3) and the radioactive precursors of macromolecular synthesis (Bachrach U and Rosenkranz HS, 1969). Results are described in Table 1. It may be seen that the synthesis of RNA, DNA and proteins was inhibited by 75%-99%.

Table 1: Inhibition of growth

Cultured cells	Control (cpm)	POX-3 (cpm)	Inhibition (%)
( <sup>3</sup> H)-uridine incorporation	73,309	770	99
( <sup>3</sup> H)-TPM (Total productive maintenance) incorporation	2,592	399	84.5
( <sup>3</sup> H)-lysine incorporation	3,153	789	75

**Viruses**

The oxidation products of polyamines are highly cytotoxic. They are anti-cancer (Averill-Bates DA, *et al.*, 2005; Bachrach U, *et al.*, 1967) and in-

hibit bacterial growth (Bachrch U and Persky S, 1964). Various antibacterial and anticancer agents are known, but our arsenal to combat viral diseases is rather limited. It was therefore of interest to find out whether polyamines can also inhibit the growth of viruses. Bacterial viruses (bacteriophages), infect bacteria such as *Escherichia coli* which grows easily. It is therefore not surprising that the phage-bacteria system served as an early model to study antiviral activities of oxidized polyamines (Figure 4).

**Antiviral Activity**

**Bacteriophages- T5**

**Plant Viruses - Tobacco Mosaic Virus**

**Human Viruses - Influenza Virus**

**Animal Viruses – Newcastle Disease Virus**

Figure 4: Antiviral activity of oxidized spermine

**Bacterial viruses**

It soon became apparent that bacteriophages differ in their susceptibility to oxidized spermine (Figure 5). Bacteriophages of the T-odd series were sensitive, while those of the T-even series were resistant (Bachrach U, *et al.*, 1963). This can be explained by the known fact, that coliphages of the T-odd series are permeable, unlike the coliphages of the T-even series. This would imply that oxidized polyamines have to penetrate in O-phage heads, to exert cytotoxicity.

	Plaque forming	Units	Permeable	
T3	1.1x10 <sup>8</sup>	9.1x10 <sup>4</sup>	4	"
T5	2.3x10 <sup>8</sup>	1.1x10 <sup>5</sup>	3	"
T7	9.6x10 <sup>8</sup>	1.5x10 <sup>3</sup>	5	"
T2	1.0x10 <sup>8</sup>	1.7x10 <sup>8</sup>	0	Non-Permeable
T4	1.7x10 <sup>8</sup>	1.1x10 <sup>8</sup>	0	"
MS2	8.0x10 <sup>8</sup>	1.0x10 <sup>4</sup>	4	RNA Phage

Figure 5: Inactivation of bacteriophages by enzymatically prepared oxidized spermine

**Anti-viral activity**

After finding that T5 coliphages are inactivated by oxidized spermine, we studied the sensitivity of different viruses to oxidized spermine. It may be seen that after exposing the viruses to oxidized spermine for 3 hours, the infectivity was reduced by log 1-4 (Table 2).

Table 2: Inactivation of viruses by oxidized spermine

Virus	Inactivation log <sub>10</sub>
Influenza virus	2
Newcastle disease	2
West Nile	4
Sindbis	4

### Plant viruses

After finding that permeable coliphages are inactivated by oxidized polyamines, plant viruses were examined. These included *Potato virus X* (PVX), *Tobacco mosaic Virus* (TMV) and *Alfafa mosaic Virus* (AMV).

When TMV viruses were treated with enzymatically-prepared oxidized spermine (100 µg/ml) for 3 hours and then rubbed on tobacco leaves (Bachrach U, *et al.*, 1965), only 5% of the control lesions were detected (Figure 6). Oxidized polyamines had no deleterious effect on plant leaves. This is a very important finding, as only a few drugs can form germ-free tobacco plants.

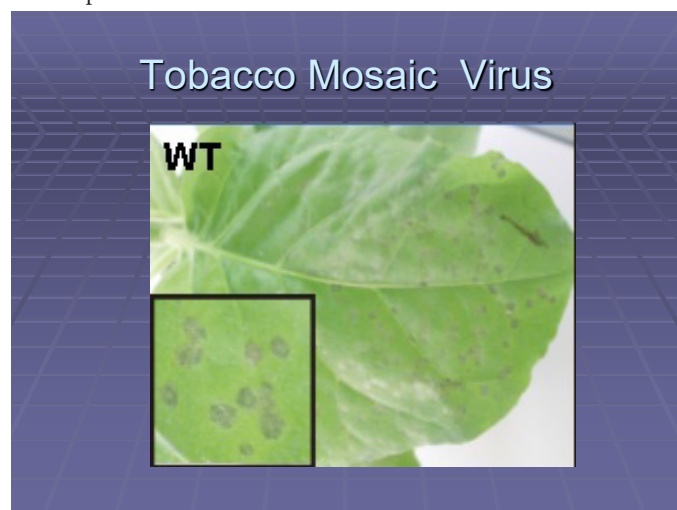


Figure 6: Inactivation of *Tobacco mosaic virus* by enzymatically prepared oxidized spermine

### Human viruses

When influenza viruses, were incubated for 24 hours with oxidized spermine (1.65 µg/ml) and then exposed to chick red blood cells, the inactivated viruses retained their property to hemagglutinate. Oxidized polyamines, interact with cellular nucleic acids and form a biological inactive complex. Viral vaccines are usually prepared by treating viruses with formalin, which interact with viral membrane proteins. Human and/or animal viruses treated with imino-dialdehydes, retain unchanged viral membranes and are therefore potent immunogens (Figure 7 and Table 3). Vaccination of animals with influenza or Newcastle disease viruses, inactivated by imino-dialdehydes, gave better results compared to conventionally prepared vaccines. This opens new possibilities to improve vaccination processes.

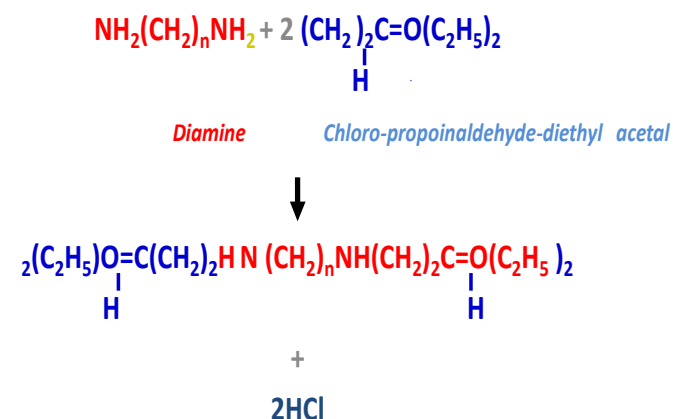


Figure 7: Synthesis of imino-aldehydes

Table 3: Immune response after injecting live or inactivated influenza virus into mice

Preparation	Antibody titer (units)
Live virus	1,280
Oxidized spermine 1.65 µmol/ml	1,280
Formaldehyde	
• 0.05%	853
• 0.03%	480
Glutaraldehyde (3.3 µmol/ml)	970

### Newcastle disease virus

Newcastle disease viruses were treated with oxidized polyamines with different structures. This included enzymatically prepared oxidized spermine and synthetic compounds containing central chains with 4 or 6 carbons. The cytotoxicity of the different compounds was compared.

It may be seen that the compound with diamino-hexane, was the most potent one and after 1 hour the viability of the viruses was reduced by 3.3 log<sub>10</sub> (Table 4). The synthetic compound with a diamino-butane chain was also very active. It may be concluded that cytotoxicities of synthetic oxidized polyamines can be modified by changing their composition and can be applied in the control of viral growth.

Table 4: Inactivation of Newcastle disease viruses by imino-aldehydes

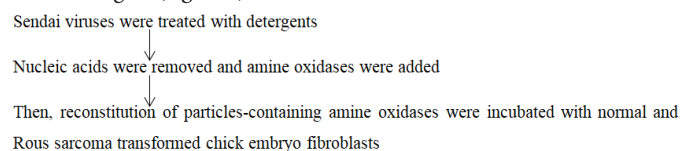
Time (hours)	Compound (0.8 mM)	P.F.U/ml	Inactivation (log <sub>10</sub> )
0	-	8.7 × 10 <sup>7</sup>	-
1	Oxidized spermine (enzymatic)	8.2 × 10 <sup>6</sup>	1
3	Oxidized spermine (enzymatic)	4.4 × 10 <sup>4</sup>	3.3
5	Oxidized spermine (enzymatic)	2.5 × 10 <sup>2</sup>	5.5
1	Oxidized spermine (synthetic)	4.7 × 10 <sup>4</sup>	3.3
3	Oxidized spermine (synthetic)	10	6
5	Oxidized spermine (synthetic)	0	7
1	Diamino-hexane	1.2 × 10 <sup>4</sup>	3.8
3	Diamino-hexane	10	6

Note: P.F.U: Plaque Forming Units  
Oxidized polyamines were incubated with viruses at 37°C for 3 hours

### Polyamines and carcinogenesis

**Gene therapy:** Cancer cells are rich in polyamines and these polycations can be oxidized by serum amine oxidase to yield cytotoxic products (Tabor CW, *et al.*, 1964). Based on these considerations, it is conceivable that cancer cells could be inactivated preferentially, as they contain polyamines, which are the substrates of amine oxidases. Normal, cells, which contain only small amounts of polyamines, should not be affected by the injected enzyme.

*Sendai viruses* can be solubilized by detergents. Removal of the detergent leads to the formation of empty reconstituted virus envelopes. When soluble macromolecules are present in the detergent-solubilized fraction, they are trapped within resealed envelopes, reconstituted after the removal of the detergent (Figure 8).



**Figure 8: Flowchart for micro-injection of amine oxides**

Bovine serum amine oxidases were trapped within reconstituted *Sendai* envelopes and they retained their activity (Table 5). *Sendai viruses* were used to inhibit the growth of cancer-bearing cells. Viruses were disintegrated by detergents and viral envelopes were separated from nuclear nuclei by centrifugation. Amine oxidases were trapped within reconstituted *Sendai viruses* and retained their activities. Reconstituted viral particles were injected into cultured fibroblasts of chick or rat embryos (Figure 9). As expected, cells rich in polyamines (transformed fibroblasts of chick or rat embryos), were more susceptible to the injected oxidases, rather than others which contain less polyamines.

When reconstituted viruses were incubated with chick embryo fibroblasts, transformed by *Rous sarcoma virus*, deformations and holes were observed in the fibroblasts. The shape of normal eukaryotic cells was not affected by reconstituted *Sendai viruses* (Bachrach U, et al., 1987). Immobilized bovine serum oxidase was also used by other investigators. The group of Agostinelli (Bachrach U, et al., 1987; Averill-Bates DA, et al., 2005) prepared poly-ethylene glycol particles containing bovine serum amine oxidase. These, were injected into mice carrying B16 tumor cells. A significant arrest of tumor growth was observed.

The group of Mondovi (Stevanato R, et al., 1989) also prepared immobilized serum amine oxidase particles.

Polyamines are a marker of proliferation and ornithine decarboxylase is an early possibly obligatory event in carcinogenesis (O'Brien TG, 1976). Interference in their activities may arrest of growth. Imino-aldehydes block the activity of polyamines (Table 3) and therefore may be regarded as a chemopreventive agent.  $\alpha$ -Difluoromethyl Ornithine (DFMO), inhibits the activity of ornithine decarboxylase and has been used to suppress the outbreak of prostate malignancies (Meyskens FI and Gerner EW, 1999). It appears that immobilized amine oxidases have a potential therapeutic value. The inert liposomes, used for trapping the enzymes have an advantage, by not being antigenic and do not trigger the production of immunological reactions. On the other hand, the reconstituted viral envelopes can be targeted and therefore be more potent and specific. Chemoprevention should be regarded as an important tool to combat cancer and should be applied widely.

**CONCLUSION**

Polyamines are oxidized by serum amine oxidase, yielding unstable cytotoxic amino-aldehydes. These compounds, inhibit the growth of bacteria, plant, animal and human viruses. Their anti-cancer activity was also reported. Stable imino-aldehydes were synthesized. They contained acetal groups, which protected aldehyde regions. The central chains of these synthetic stable compounds contained 4 or 6 carbons. They were very active and inhibited the growth of influenza viruses and may be the basis for improved vaccination processes. Serum amine oxidases were trapped within reconstituted *Sendai virus* envelopes and injected into normal and transformed fibroblasts. The growth of cancer cells was preferentially inhibited; this may be defined as an example of "gene therapy". The synthetic imino-aldehydes also prevented carcinogenesis and may play a role as a chemopreventive agent.

It may be concluded that polyamines and their biologically and synthetic derivatives are biologically active and may contribute to the improvement of vaccination, gene therapy and chemoprevention.

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**Table 5: Effect of microinjected amines and diamine oxidases on the ultrastructure of eukaryotic cultured cells**

Culture	Incorporation (ct/min/culture dish)	
	Thymidine	Leucine
Normal cells	11,500	12,000
Normal cells+microinjected oxidases	10,800	10,000
Transformed cells	10,125	10,000
Transformed cells+microinjected oxidases	3,800	3,200



**Figure 9: Scanning electron micrograph of transformed chick embryo fibroblasts treated with reconstituted envelopes containing amine oxidase**

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