## Notch Pathway and its Role in Cardiovascular System: Review

Amira M. Badr<sup>1,2</sup> and Yasmen F. Mahran<sup>3,4</sup>

<sup>1</sup>Pharmacology and Toxicology Department, Faculty of Pharmacy, King Saud University, Riyadh, KSA <sup>2</sup> Pharmacology and Toxicology Department, Faculty of Pharmacy, Ain Shams University, Cairo <sup>3</sup>Department of Pharmaceutical Sciences, Faculty of Pharmacy, Princess Neurab hist Abdulashman University, Pharmaceutical Sciences, Faculty of Pharmacy, Princess Neurab hist Abdulashman University, Pharmaceutical Sciences, Faculty of Pharmacy, Princess Neurab hist Abdulashman University, Pharmaceutical Sciences, Faculty of Pharmacy, Princess Neurab hist Abdulashman University, Pharmaceutical Sciences, Faculty of Pharmaceutical Sciences, Faculty of Pharmacy, Princess Neurab hist Abdulashman University, Pharmaceutical Sciences, Faculty of Pharmaceutical Sciences, Pharmaceu

<sup>3</sup>Department of Pharmaceutical Sciences, Faculty of Pharmacy, Princess Nourah bint Abdulrahman University, Riyadh, KSA. <sup>4</sup>Faculty of Pharmacy, Ain Shams University, Cairo, Egypt.

## \*Corresponding author: Yasmen F. Mahran

Department of Pharmaceutical Sciences, Faculty of Pharmacy, Princess Nourah bint Abdulrahman University, Riyadh, KSA **Email :** Jassie\_81@hotmail.com; yfmahranr@pnu.edu.sa

#### ABSTRACT

The Notch signaling pathway is a core signal transduction mechanism that control diverse cellular processes. Its activity is mandatory for normal embryonic development, as it control the transcription of genes implicated in cellular proliferation, differentiation, migration, and apoptosis. It is considered as a key regulator of neonatal cardiomyocyte development; thus, its abnormality is strongly correlated to cardiovascular congenital malformations. Despite, Notch pathway is considered relatively absent in adult heart; recent studies included evidence of the upregulation of Notch signaling pathway in response to some cardiac insults. In our review, we briefly discuss the possible role of Notch Pathway in adult cardiomyocytes and other cells involved in the cardiovascular health, and its possible contribution to cardiovascular diseases.

### **INTRODUCTION**

Recently, different studies reports that Notch pathway is being activated in injured stressed hearts, such as in coronary heart disease (1) and hypoxia/ reoxygenation (2). Contradictory results are also available considering the outcomes of Notch pathway upregulation in adult heart, whether it is cardioprotective (3), or may be one of the mechanisms that contributes to cardiac pathogenesis (4,5). This may be explained by the cell context in which Notch Pathway is upregulated, the diverse number of Notch receptors and ligands that confer different responses, the existence of both canonical and noncanonical activation mechanisms, and the widespread of different Notch receptors in different cells that are involved in cardiovascular pathogenesis; cardiomyocytes, endothelial cells, and vascular smooth muscle cells (VSMC) (6,7). Moreover, considering the role of Notch signaling in promoting inflammation and regulation of macrophages' activation and proliferation; which also contribute to the progression of considerable number of cardiovascular diseases; may help to further understand the cardiovascular influences of Notch Pathway activation (8,9).

### The Notch Signaling Pathway

The Notch Pathway is an evolutionally pathway, that plays a well-defined key role during embryonic life, and in specific tissues in adults. However, in many tissues it is relatively absent in adults, and is being activated in response to specific insults (10). In the kidney, for example, the Notch pathway is activated during nephrgenesis, being inhibited in the neonatal and adult stages, and it is reactivated in situations of kidney damage and its activation correlated with the degree of renal damage (11). It is a fundamental pathway that seems to have a key role in many vital processes such as neurogenesis, gastric mucosal cells' proliferation, angiogenesis, and hematopoiesis (12-16), and its dysregulation is correlated to tissue abnormalities such as fibrogenesis and carcinogenesis (17,18). Obviously, it helps to determine cell fate, as it controls the transcription

**Keywords:** Notch pathway, cardiovascular diseases, Notch receptors, Canonical Notch Activation

#### Correspondence:

Yasmen F. Mahran Department of Pharmaceutical Sciences, Faculty of Pharmacy, Princess Nourah bint Abdulrahman University, Riyadh, KSA Email : Jassie\_81@hotmail.com; yfmahranr@pnu.edu.sa

of genes involved in cellular proliferation and differentiation, as well as genes involved in fibrosis and apoptosis (12,19). The ultimate effect will depend largely on the cell context in which Notch Pathway is activated (17).

### **Receptors and Ligands**

Up till now, a family of four receptors (Notch 1, 2, 3, and 4) and five ligands (Jagged 1, Jagged 2, and Delta-like-1, 3, and 4) have been identified in humans and mice. Canonical and noncanonical identified Ligands have been known for either agonistic or antagonistic signaling , which in turn might affect Notch signaling(20). Receptors and ligands are expressed on the cell surface and regulate adjacent cell-cell communication. They are composed of polypeptide chains with extracellular, transmembrane and functional intracellular domain (NICD). Both extracellular domain (NECD) and the single polypeptide chain of transmembrane fragment (TMF) are the polypeptides which encounter for the mature membrane-bound form of Notch (21,22).

### The Intracellular Cascade

Notch signaling can be activated via either canonically (more common) or noncanonically (17). During canonical Notch Pathway activation, the Notch receptors; that were synthesized in the endoplasmic reticulum and the Golgi apparatus as a single-chain; undergoes modification by glycosylation and proteolysis at S1, to produce the mature receptor that is being expressed on cell membrane (23). Notch signaling starts upon coupling of ligand and receptor coupling; expressed on the surface of two opposing cells; followed by endocytosis of the complex by ligand expressing cell. Subsequently, the receptor chain will be further cleaved by two more proteases at S2 and S3 leads to cleavage at the plasma membrane by two other proteases at S2 and S3 that removes the extracellular fragment of the heterodimer. Extracellular S2 cleavage is due to the tumor necrosis factor-alpha converting enzyme (TACE), a disintegrin and metalloproteinase domaincontaining protein 10 (ADAM10), and (ADAM17). S3 cleavage is achieved by a  $\gamma$ -secretase, with the ultimate

release of the active NICD into the cytoplasm (Figure 1) (24).

NICD is a transcriptional regulator, it translocates to the nucleus and forms a complex with one of CBF1/Su(H)/Lag-1 (CSL) members; the recombinant signal binding protein 1 for Jk (RBPjk). In the case of the absence of Notch activation, CSL represses transcription of Notch target genes by recruiting corepressor proteins to form a multiprotein transcriptional repressor complex (25). NICD binding to CSL displaces corepressors proteins bound to CSL, and permits the binding of the Mastermind like protein (MAML); a transcriptional coactivator; to the

complex, which thus culminates in activated expression of Notch specific genes (26) (Figure 2).

The most prominent Notch activated genes are the hairy and enhancer of split (Hes) and the hairy and enhancer of the split with YRPW motif (Hey) gene families, which are transcription regulators. Hes genes are believed to play important role in neural and endocrine functions, however, Hey genes are more involved in the cardiovascular system (27). Other Notch target genes include cyclin D1 (implicated in cell cycle progression), Bcl-2 (inhibitors of apoptosis), cyclin A, and Nuclear factor -kB (NF-kB) (28,29) (Figure 2).



Figure 1: Activation of The Notch Receptor by Notch Ligand from A neighboring Cell.

Notch ligand is are located on the cell surface of signaling cell. The Notch receptor is located on the surface of the signal-receiving cell. Upon ligand binding, the receptor chin is processed at the S1 site by a furin protease, followed by 2 consecutive cleavages at S2 and S3, by ADAM

and  $\gamma$ -secretase, respectively. Those cleavages free the NICD that translocates to the nucleus. and binds to CSL **(30)**. ADAM: A Disintegrin and Metalloprotease-containing protease, NICD: The Notch intracellular domain



Figure 2: A Model of Canonical Notch Activation.

In the nucleus, NICD binds to (RBP-j) one of CSL members, the binding of NICD results in displacement of transcriptional repressors (Co-R) by coactivators (Co-A), in presence of the transcriptional coactivator Mastermind (MAML). The results is activation of transcription of Notch target genes (e.g., Hes1, Hes5) (31).

### Noncanonical Notch Signaling Pathways

In addition to canonical activation of Notch Pathway, it was found that Notch can also be activated by a wide variety of noncanonical ligands; this may explain the variable and missing information about Notch activation outcomes (30).

This pathway is even variable, including Notch pathway effects that progress independently of CSL, without S3 cleavage, those initiated without ligand binding, or signals that activate CSL-genes without NICD release. Among suggested mechanisms of noncanonical Notch signaling are induction of non-CSL transcriptional factors, including  $\beta$ -catenin, hypoxia-inducible factor-1 $\alpha$ , NF- $\kappa$ B, and estrogen receptor (ERα). For example, Notch can activate through R-Ras to promote cell adhesion. Alternatively, Notch may interact with IKKa in the NF-kB pathway or lymphoid enhancer-binding factor 1 in the Wnt pathway. Collectively, our knowledge about the role of the noncanonical Notch signaling in biology and pathophysiology remains primitive (Figure 3) (28).

# The effect of Notch ligands and feedback loop on Notch signaling activity

From the above mentioned, it had been reported that the pleiotropic nature of Notch pathway has been accounted by the canonical and non-canonical Notch ligands, transmembrane proteins, which might act as agonists or antagonists through a specific molecular transduction mechanisms (20). With respect to the canonical DSL (Delta, Serrate, Lag2) ligands, the nature of the ligands might affect being either amplifying or repressing the Notch signaling. Parks and coworkers identified the structure-function analysis of these ligands and found that

the DSL domain together with the flanking N-terminal (NT) domain and first two Epidermal Growth Factor (EGF) repeats are required for DSL ligands to bind Notch (32). DSL ligands can either activate or inhibit the Notch signaling depending on the nature of cell surface interaction (20). For instance, a Trans-interaction between the DSL ligand on one cell (signal-sending cell) and Notch receptor on the opposing cell (signal-receiving cell) is required to amplify the Notch signaling (32). On the other hand, DSL ligand can inhibit the Notch signaling when interacts with the Notch receptor within the same cell, Cis -interactions (33,34). In addition, DSL ligands activity are also regulated by glycosylation of Notch through modulating the ligand-binding properties (35). Nevertheless, the molecular basis of cis-interaction and the exact mechanism by which it could affect Notch are not fully understood. In addition to the cis- and transinteractions, it had been thought by some that Notch ligands might have a concentration dependent inhibitory effect on Notch signaling activity. High levels of ligand causes an inhibitory effect while low levels activate the Notch signaling (36)

Another molecular explanation for the pleiotropic nature of Notch signaling is the non-canonical Notch ligands such as Delta-like 1 (Dlk-1) (37). Indeed, Dlk-1 can only support cis-interaction with the Notch resulting in cis-inhibition and not trans-activation of Notch signaling (20,38).

Besides the Notch ligands modulatory effects, Notchmediated lateral inhibition is one mechanism that control the fate of two adjacent cells into opposite fates, amplification of suppression, although having the same differentiation (39,40). One explanation is that suppression of Notch signaling occurred in the winner cell, which produces more Delta ligands while amplification of Notch signaling is usually happened in the loser cell which downregulates Delta production. Nevertheless, the mechanisms that activate the Notch signaling in the loser cell are the same which terminates Notch signaling in the winner cell (39,41).

# The Role of Notch Pathway in the Cardiovascular System

The Notch pathway plays a magnificent role in the cardiovascular system. It is a key regulator in multiple processes during embryonic cardiovascular system development and maintenance (41), such as the proliferation and differentiation of vascular cells and

cardiomyocytes. Sequentially, Notch dysregulation is believed to contribute largely to congenital cardiovascular diseases and strong evidence grow about a possible role of Notch in the etiology of many cardiovascular complications in adults, as well (42). However, like many other signaling pathways, its effect is complexed interactions with other pathways, in addition to being itself including many different receptors and ligands, which makes its effect difficult to be predicted (43).



Figure 3: Notch canonical versus non-canonical signaling.

Canonical Notch pathway is illustrated on the left side of the figure. Notch receptor is activated by ligand on a neighboring cell, with subsequent proteolytic cleavages releasing the NICD. The non-canonical signaling pathway is illustrated on the right side of the figure and it illustrates that non-canonical Notch activation may occur with or without ligand, with or without receptor cleavage. In Noncanonical Notch signaling is independent of CSL, and interaction with many different pathways can occur, either in the cytoplasm or in the nucleus. Abbreviations: ADAM, a disintegrin and metalloprotease; NEXT, Notch extracellular truncation; NICD, Notch intracellular domain; Co-R, corepressor; Co-A, coactivator **(17)**.

### The Role of Notch Pathway in Cardiomyocytes

Cardiomyocytes' stem cell precursors express considerable levels of Notch1 receptor, and Notch activation is mandatory for their proliferation. But the role of the Notch pathway in cardiomyocytes after birth is still not clear. Rat neonatal cardiomyocytes were found to actively proliferate, and their proliferation is correlated with high Notch expression. However, those cells then lose their proliferative power, correlated also with negligible Notch expression (41,44).

Recently, evidence of the reactivation of the Notch pathway in response to cardiac injury has been elucidated; e.g. the expression of Notch receptors has been detected in myocardial biopsies of patients with heart failure (45). However, the specific role and impact of Notch reactivation are still not well characterized. Studies showed that Notch 1 is reactivated in cardiomyocytes in ischemic heart and post myocardial infarction in adult rats, and it inhibited cardiomyocytes apoptosis; probably by activating the Akt pathway. These findings suggest that Notch pathway may reactivate in adult cardiomyocytes, and aids in cardiomyocytes conservation (3). Kratsios's results were also in agreement with this postulation, as he found that transgenic mice overexpressing Jagged1 (a Notch ligand) on cardiomyocytes showed better cardiac performance and less fibrosis as compared to wild animals (46). Notch mediated cardioprotection can be explained in terms of increasing cardiomyocyte survival, antiapoptotic effects, promoting angiogenesis, and stimulation cardiac stem cells proliferation (41).

Contrarily, Campa *et al.* have shown that activation of Notch in mature cardiomyocytes is correlated with apoptosis and concluded that prolonged Notch activation can thus contribute to cardiac pathogenesis (47). Similarly, Zheng *et al.* concluded that elevated expression of Notch1 was positively correlated to that of NF- $\kappa$ B in coronary heart disease rat cardiomyocytes, and that Notch1 activation plays a role in the development of coronary heart disease (48). So it is believed that the effect of active Notch signaling in adult cardiomyocytes will largely depend on the timing and dosing of the activation, thus whether Notch pathway will induce cardiomyocytes apoptosis and therefore cardiotoxicity, or Notch will be unregulated in advance to protect cardiomyocytes against toxic insults (47,48).

# The Role of Notch Signaling in Vascular Endothelial Cells

The Notch signaling is an important regulator of endothelial cell proliferation. It works in harmony and cooperates with different pathways to orchestrate the function of endothelial cells. Therefore, Notch receptors; 1, 2, and 4; as well as its ligands; Jagged 1 and 2 and DLL 1 and 4 are normally expressed in the endothelium of adults (41). It has been shown that Notch activating induces endothelial cell proliferation (49). Moreover, It also contributes to maintaining the integrity of the endothelium in stressful conditions such as ischemia and inflammation, mechanism may include antiapoptotic activity via increasing Bcl-2, in addition to modulating angiogenesis (50,51). Accordingly, it has been demonstrated that Notch inhibition leads to the induction of endothelial cells apoptosis and non-functional angiogenesis (52). The Notch endothelial protective activity was more imminent in cardiac allograft vessels with dysfunctional Notch 4 expression, in which endothelial dysfunction was prominent (50). Moreover, Notch signaling, by controlling the function of endothelial cells and maintaining endothelium integrity, plays a significant role in the formation of atherosclerotic plaques. Notch inhibition was found to worsen atherosclerosis by enhancing endothelial cells dysfunctions (41).

However, on the other hand, Liu *et al.* reported that Notch signaling regulated genes to be significantly upregulated in atherosclerotic loci (53). Similarly, Nus *et al.* found a correlation between endothelial Jag1–RBPJ complex and inflammation, via upregulating NF- $\kappa$ B and vascular cell adhesion molecule (54). Also, Quillard *et al.* have shown that increased Notch activity was associated with increased endothelial cells apoptosis, in response to inflammation (50). These contradictory results can be explained by the wide variability of receptors and ligands,

and both the canonical and non-canonical signaling mechanisms (55).

## The Role of Notch Pathway in the Vascular Smooth Muscle Cells

Notch pathway was also found to be a key regulator maintaining vascular smooth muscle cells (VSMCs) structure and function. The VSMCs expresses Notch receptors; 2 and 3; and jag 1 as the main ligand), and Notch 2 and Notch 3 mutations was found to induce defects in VSMCs development, giving a strong evidence of the implication of Notch pathway in the regulation of VSMCs differentiation, maturation, and recruitment during angiogenesis. It is also believed that Notch receptors mediate endothelial cells to VSMCs communication, and affect the phenotype of VSMCs (27,41,55,56). Experiments have shown that activation of Notch signaling not only increases proliferation but also prevents apoptosis of VSMCs. Recently, the Notch pathway has been strongly correlated to the pathogenesis of pulmonary hypertension and cerebral autosomal dominant arteriopathy via affecting VSMCs (43,57).

### The Role of Notch Signaling in Macrophages

Although macrophages are not part of the cardiovascular system, but it is well-established that macrophages play an important role in cardiovascular pathogenesis, e.g. atherosclerosis and ischemia. M1 macrophages exaggerate post-myocardial infarction injury bv inducing inflammation and scaring, while the anti-inflammatory M2 macrophages infiltration promotes tissue repair, thus the ratio of M1/M2 is of great importance (58). Notch pathway plays a significant role in inflammation by mainly affecting macrophages activity. Macrophages are divided into highinflammatory M1; secreting proinflammatory cytokines such as IL-6 and TNF- $\alpha$ ; and an anti-inflammatory M2; secreting anti-inflammatory cytokines such as IL-4 and IL-10 (59,60). It was reported that Notch1 induces the differentiation of M1 macrophages, increasing the M1/M2 ratio, and induces inflammation by increasing synthesis of IL-6 and TNF-α. Accordingly, Notch1 inhibition culminates in reduced M1/M2 ration, with increased secretion of Il-4 and IL-10 (Figure 4) (5,6,61,62).



Figure 4: Notch Pathway in Inflammation.

Notch signaling affects inflammation by M1 macrophage activation via NF- $\kappa$ B. M1 secretes TNF- $\alpha$  and IL-1 $\beta$  that promotes inflammation. On the other hand, Notch activation inhibits M2 polarization this reducing IL-10, that inhibits inflammatory process (63).

In addition to affecting macrophage differentiation, Notch is also implicated in their migration and secretion of inflammatory mediators. Aoyama *et al.* concluded that Notch inhibition reduces macrophages migration and infiltration into atherosclerotic plaques. Other studies showed that proinflammatory mediators, such as lipopolysaccharides and IL-1 $\beta$ , induce the expression of Notch Dll4 receptor in macrophages. Dll4 induces further release of proinflammatory cytokines, thereby initiating a positive feedback loop. *Besides*, incubation of macrophages with Dll4 stimulates the transcription of proinflammatory genes, e.g., iNOS, and activates Akt, and NF- $\kappa$ B. Collectively, inhibition of canonical Notch Pathway is followed by inhibition of macrophages inflammatory responses (64,65).

In the heart, Yin *et al.* found that M1 macrophages infiltrating the infarct area express mainly Notch 1, and their infiltration was inhibited by the use of a Notch inhibitor, associated with increased M2 macrophages and better cardiac function following myocardial infarction (9). It was also found that DLL 4 accelerate the inflammation of the heart vessels, and that antibody to DLL 4 suppresses M1 macrophages (66). That from these data, it can be concluded that Notch activation in macrophages induces inflammation, mainly by increasing M1/M2 ratio, with DII4 and Notch1 are considered the main keyplayers (67).

### CONCLUSION

There is a growing evidence regarding the implication of Notch Pathway in cardiovascular diseases' development. Better understanding of the diversity and contradictory effects of Notch signaling needs further research, as it may help in ameliorating or even preventing the development of different cardiac insults.

### ACKNOWLEDGEMENT

This research was funded by the Deanship of Scientific Research at Princess Nourah bint Abdulrahman University through the Fast-track Research Funding Program.

### **CONFLICT OF INTEREST**

Authors declare no conflict of interest.

### REFERENCES

- Zheng J, Zhang S, Wang T. Expression of Notch-1 and nuclear factor-κB signal pathway in myocardial cells of coronary heart disease rats. Exp Ther Med. 2019 Mar;17(3):1587–92.
- Zhu P, Yang M, He H, Kuang Z, Liang M, Lin A, et al. Curcumin attenuates hypoxia/reoxygenation-induced cardiomyocyte injury by downregulating Notch signaling. Molecular Medicine Reports [Internet]. 2019 Aug 1 [cited 2019 Dec 7];20(2):1541–50. Available from: http://www.spandidospublications.com/10.3892/m mr.2019.10371/abstract
- 3. Gude NA, Emmanuel G, Wu W, Cottage CT, Fischer K, Quijada P, *et al.* Activation of Notch-mediated protective signaling in the myocardium. Circulation Research. 2008.

- Pang X-F, Lin X, Du J-J, Zeng D-Y. Downregulation of microRNA-592 protects mice from hypoplastic heart and congenital heart disease by inhibition of the Notch signaling pathway through upregulating KCTD10. J Cell Physiol. 2019 May;234(5):6033–41.
- 5. Wang X, Yu Y. MiR-146b protect against sepsis induced mice myocardial injury through inhibition of Notch1. J Mol Histol. 2018 Aug;49(4):411–7.
- Singla RD, Wang J, Singla DK. Regulation of Notch 1 signaling in THP-1 cells enhances M2 macrophage differentiation. Am J Physiol Heart Circ Physiol. 2014 Dec 1;307(11):H1634-1642.
- Falo-Sanjuan J, Bray SJ. Decoding the Notch signal [Internet]. Vol. 62, Development, Growth & Differentiation. John Wiley & Sons, Ltd; 2020 [cited 2020 Jun 29]. p. 4–14. Available from: https://onlinelibrary.wiley.com/doi/abs/10.1111/d gd.12644
- 8. Ilinykh A, Pinto AR. The Role of Cardiac Tissue Macrophages in Homeostasis and Disease. Adv Exp Med Biol. 2017; 1003:105–18.
- Yin J, Hu H, Li X, Xue M, Cheng W, Wang Y, *et al.* Inhibition of Notch signaling pathway attenuates sympathetic hyperinnervation together with the augmentation of M2 macrophages in rat's postmyocardial infarction. Am J Physiol, Cell Physiol. 2016 Jan 1;310(1):C41-53.
- Perrimon N, Pitsouli C, Shilo B-Z. Signaling mechanisms controlling cell fate and embryonic patterning. Cold Spring Harbor perspectives in biology. 2012;4(8): a005975.
- 11. Marquez-Exposito L, Cantero-Navarro E, Lavoz C, Fierro-Fernández M, Poveda J, Rayego-Mateos S, *et al.* Could the Notch signaling pathway be a potential therapeutic option in renal diseases? Nefrología (English Edition). 2018;38(5):466–475.
- 12. Reichrath J, Reichrath S. Notch Pathway and Inherited Diseases: Challenge and Promise. Adv Exp Med Biol. 2020; 1218:159–87.
- van Es JH, van Gijn ME, Riccio O, van den Born M, Vooijs M, Begthel H, *et al.* Notch/γ-secretase inhibition turns proliferative cells in intestinal crypts and adenomas into goblet cells. Nature [Internet]. 2005 Jun 1;435(7044):959–63. Available from: https://doi.org/10.1038/nature03659
- Kofler NM, Shawber CJ, Kangsamaksin T, Reed HO, Galatioto J, Kitajewski J. Notch Signaling in Developmental and Tumor Angiogenesis. Genes Cancer [Internet]. 2011 Dec [cited 2020 Jun 29];2(12):1106–16. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC34 11124/
- Engler A, Zhang R, Taylor V. Notch and Neurogenesis. In: Borggrefe T, Giaimo BD, editors. Molecular Mechanisms of Notch Signaling [Internet]. Cham: Springer International Publishing; 2018 [cited 2020 Jun 29]. p. 223–34. (Advances in Experimental Medicine and Biology). Available from: https://doi.org/10.1007/978-3-319-89512-3\_11
- Bigas A, Espinosa L. Hematopoietic stem cells: to be or Notch to be. Blood [Internet]. 2012 Apr 5 [cited 2020 Jun 29];119(14):3226–35. Available from: https://ashpublications.org/blood/article/119/14/ 3226/29567/Hematopoietic-stem-cells-to-be-or-Notch-to-be

- 17. D'Souza B, Meloty-Kapella L, Weinmaster G. Canonical and non-canonical Notch ligands. In: Current topics in developmental biology. Elsevier; 2010. p. 73–129.
- Zhu C, Kim K, Wang X, Bartolome A, Salomao M, Dongiovanni P, *et al.* Hepatocyte Notch activation induces liver fibrosis in nonalcoholic steatohepatitis. Sci Transl Med. 2018 21;10(468).
- Hu B, Phan SH. Notch in fibrosis and as a target of antifibrotic therapy. Pharmacological Research [Internet]. 2016 Jun 1 [cited 2020 Jun 29]; 108:57–64. Available from: http://www.sciencedirect.com/science/article/pii/S 1043661816303097
- D'souza B, Miyamoto A, Weinmaster G. The many facets of Notch ligands. Oncogene. 2008 Sep 1;27(38):5148–67.
- Hossain F, Sorrentino C, Ucar DA, Peng Y, Matossian M, Wyczechowska D, *et al.* Notch Signaling Regulates Mitochondrial Metabolism and NF-κB Activity in Triple-Negative Breast Cancer Cells via IKKα-Dependent Non-canonical Pathways. Front Oncol. 2018; 8:575.
- 22. Steinbuck MP, Winandy S. A Review of Notch Processing with New Insights into Ligand-Independent Notch Signaling in T-Cells. Front Immunol [Internet]. 2018 [cited 2020 Oct 30];9. Available from: https://www.frontiersin.org/articles/10.3389/fimm u.2018.01230/full
- 23. Guo H, Lu Y, Wang J, Liu X, Keller ET, Liu Q, *et al.* Targeting the Notch signaling pathway in cancer therapeutics. Thorac Cancer. 2014 Nov;5(6):473–86.
- Kandachar V, Roegiers F. Endocytosis and control of Notch signaling. Curr Opin Cell Biol. 2012 Aug;24(4):534–40.
- 25. Borggrefe T, Oswald F. The Notch signaling pathway: transcriptional regulation at Notch target genes. Cell Mol Life Sci. 2009 May;66(10):1631–46.
- Kopan R, Ilagan MXG. The canonical Notch signaling pathway: unfolding the activation mechanism. Cell. 2009;137(2):216–233.
- 27. Boucher JM, Gridley T, Liaw L. Molecular pathways of notch signaling in vascular smooth muscle cells. Frontiers in physiology. 2012; 3:81.
- Espinoza I, Miele L. Notch inhibitors for cancer treatment. Pharmacol Ther. 2013 Aug;139(2):95– 110.
- 29. Huang Q, Li J, Zheng J, Wei A. The Carcinogenic Role of the Notch Signaling Pathway in the Development of Hepatocellular Carcinoma. J Cancer. 2019 Feb 26;10(6):1570–9.
- Andersen P, Uosaki H, Shenje LT, Kwon C. Noncanonical Notch signaling emerging role and mechanism. Trends in Cell Biology. 2012 May;22(5):257–65.
- Arai MA, Akamine R, Tsuchiya A, Yoneyama T, Koyano T, Kowithayakorn T, *et al.* The Notch inhibitor cowanin accelerates nicastrin degradation. Sci Rep. 2018 29;8(1):5376.
- 32. Nichols JT, Miyamoto A, Weinmaster G. Notch signaling--constantly on the move. Traffic. 2007 Aug;8(8):959–69.
- Glittenberg M, Pitsouli C, Garvey C, Delidakis C, Bray S. Role of conserved intracellular motifs in Serrate signalling, cis-inhibition and endocytosis. EMBO J. 2006 Oct 18;25(20):4697–706.

- 34. Fiúza U-M, Arias AM. Cell and molecular biology of Notch. J Endocrinol. 2007 Sep;194(3):459–74.
- 35. Irvine KD. A notch sweeter. Cell. 2008 Jan 25;132(2):177–9.
- Klein T, Arias AM. Interactions among Delta, Serrate and Fringe modulate Notch activity during Drosophila wing development. Development. 1998 Aug 1;125(15):2951–62.
- Bachmann E, Krogh TN, Højrup P, Skjødt K, Teisner B. Mouse fetal antigen 1 (mFA1), the circulating gene product of mdlk, pref-1 and SCP-1: isolation, characterization and biology. J Reprod Fertil. 1996 Jul;107(2):279–85.
- Baladrón V, Ruiz-Hidalgo MJ, Nueda ML, Díaz-Guerra MJM, García-Ramírez JJ, Bonvini E, *et al.* dlk acts as a negative regulator of Notch1 activation through interactions with specific EGF-like repeats. Exp Cell Res. 2005 Feb 15;303(2):343–59.
- Sancho R, Cremona CA, Behrens A. Stem cell and progenitor fate in the mammalian intestine: Notch and lateral inhibition in homeostasis and disease. EMBO Rep. 2015 May;16(5):571–81.
- Lemmon MA, Freed DM, Schlessinger J, Kiyatkin A. The Dark Side of Cell Signaling: Positive Roles for Negative Regulators. Cell. 2016 Mar 10;164(6):1172– 84.
- 41. Fior R, Henrique D. "Notch-Off": a perspective on the termination of Notch signalling. Int J Dev Biol. 2009;53(8–10):1379–84.
- Aquila G, Pannella M, Morelli MB, Caliceti C, Fortini C, Rizzo P, et al. The role of Notch pathway in cardiovascular diseases. Glob Cardiol Sci Pract [Internet]. 2013 Dec 30 [cited 2019 Nov 8];2013(4):364–71. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC39 91209/
- Luxán G, D'Amato G, MacGrogan D, de la Pompa JL. Endocardial Notch Signaling in Cardiac Development and Disease. Circ Res. 2016 Jan 8;118(1): e1–18.
- 44. Baeten JT, Lilly B. Notch Signaling in Vascular Smooth Muscle Cells. Adv Pharmacol. 2017; 78:351–82.
- 45. Zheng Y-H, Li F-D, Tian C, Ren H-L, Du J, Li H-H. Notch γ-Secretase Inhibitor Dibenzazepine Attenuates Angiotensin II-Induced Abdominal Aortic Aneurysm in ApoE Knockout Mice by Multiple Mechanisms. PLoS One [Internet]. 2013 Dec 16 [cited 2019 Dec 10];8(12). Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC38 65307/
- Øie E, Sandberg WJ, Ahmed MS, Yndestad A, Lærum OD, Attramadal H, *et al*. Activation of Notch signaling in cardiomyocytes during post-infarction remodeling. Scand Cardiovasc J. 2010 Dec;44(6):359–66.
- 47. Kratsios P, Catela C, Salimova E, Huth M, Berno V, Rosenthal N, *et al.* Distinct roles for cell-autonomous Notch signaling in cardiomyocytes of the embryonic and adult heart. Circ Res. 2010 Feb 19;106(3):559– 72.
- 48. Campa VM, Gutiérrez-Lanza R, Cerignoli F, Díaz-Trelles R, Nelson B, Tsuji T, *et al.* Notch activates cell cycle reentry and progression in quiescent cardiomyocytes. J Cell Biol. 2008 Oct 6;183(1):129– 41.
- Zheng J, Zhang S, Wang T. Expression of Notch-1 and nuclear factor-κB signal pathway in myocardial cells of coronary heart disease rats. Experimental and Therapeutic Medicine. 2019 Mar 1;17(3):1587–92.

- 50. Jiang H, Cheng XW, Shi G-P, Hu L, Inoue A, Yamamura Y, *et al.* Cathepsin K-mediated Notch1 activation contributes to neovascularization in response to hypoxia. Nat Commun. 2014 Jun 4; 5:3838.
- 51. Quillard T, Coupel S, Coulon F, Fitau J, Chatelais M, Cuturi MC, *et al.* Impaired Notch4 activity elicits endothelial cell activation and apoptosis: implication for transplant arteriosclerosis. Arterioscler Thromb Vasc Biol. 2008 Dec;28(12):2258–65.
- 52. Rizzo P, Miele L, Ferrari R. The Notch pathway: a crossroad between the life and death of the endothelium. Eur Heart J [Internet]. 2013 Aug 21 [cited 2019 Dec 9];34(32):2504–9. Available from: https://academic.oup.com/eurheartj/article/34/32 /2504/632209
- 53. Xue S, He L, Zhang X, Zhou J, Li F, Wang X. Expression of Jagged1/Notch3 Signaling Pathway and their Relationship with the Tumor Angiogenesis in TNBC. Arch Med Res. 2017 Feb;48(2):169–79.
- 54. Liu Z-J, Tan Y, Beecham GW, Seo DM, Tian R, Li Y, et al. Notch Activation Induces Endothelial Cell Senescence and Pro-inflammatory Response: Implication of Notch Signaling in Atherosclerosis. Atherosclerosis [Internet]. 2012 Dec [cited 2019 Nov 23];225(2):296–303. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC35 02717/
- 55. Nus M, Martínez-Poveda B, MacGrogan D, Chevre R, D'Amato G, Sbroggio M, *et al.* Endothelial Jag1-RBPJ signalling promotes inflammatory leucocyte recruitment and atherosclerosis. Cardiovasc Res. 2016 Nov 1;112(2):568–80.
- 56. Mack JJ, Iruela-Arispe ML. NOTCH regulation of the endothelial cell phenotype. Curr Opin Hematol [Internet]. 2018 May [cited 2019 Nov 8];25(3):212–8. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC59 02133/
- 57. Kuhnert F, Kirshner JR, Thurston G. Dll4-Notch signaling as a therapeutic target in tumor angiogenesis. Vascular Cell. 2011 Sep 18;3(1):20.
- Geraci MW, Moore M, Gesell T, Yeager ME, Alger L, Golpon H, *et al.* Gene expression patterns in the lungs of patients with primary pulmonary hypertension: a gene microarray analysis. Circ Res. 2001 Mar 30;88(6):555–62.
- 59. ter Horst EN, Hakimzadeh N, van der Laan AM, Krijnen PAJ, Niessen HWM, Piek JJ. Modulators of Macrophage Polarization Influence Healing of the Infarcted Myocardium. Int J Mol Sci. 2015 Dec 10;16(12):29583–91.
- Atri C, Guerfali FZ, Laouini D. Role of Human Macrophage Polarization in Inflammation during Infectious Diseases. Int J Mol Sci [Internet]. 2018 Jun 19 [cited 2019 Dec 14];19(6). Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC60 32107/
- Tabas I, Lichtman AH. Monocyte-Macrophages and T Cells in Atherosclerosis. Immunity. 2017 17;47(4):621–34.
- 62. Singla DK, Johnson TA, Tavakoli Dargani Z. Exosome Treatment Enhances Anti-Inflammatory M2 Macrophages and Reduces Inflammation-Induced Pyroptosis in Doxorubicin-Induced Cardiomyopathy. Cells [Internet]. 2019 Oct 9 [cited 2019 Dec 8];8(10). Available from:

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC68 30113/

- 63. Singla DK, Wang J, Singla R. Primary human monocytes differentiate into M2 macrophages and involve Notch-1 pathway. Can J Physiol Pharmacol. 2017 Mar;95(3):288–94.
- Bi P, Kuang S. Notch signaling as a novel regulator of metabolism. Trends in Endocrinology & Metabolism. 2015 May 1;26(5):248–55.
- 65. Aoyama T, Takeshita K, Kikuchi R, Yamamoto K, Cheng XW, Liao JK, *et al*. γ-Secretase inhibitor reduces diet-induced atherosclerosis in apolipoprotein Edeficient mice. Biochem Biophys Res Commun. 2009 May 29;383(2):216–21.
- Fung E, Tang S-MT, Canner JP, Morishige K, Arboleda-Velasquez JF, Cardoso AA, *et al.* Delta-like 4 induces notch signaling in macrophages: implications for inflammation. Circulation. 2007 Jun 12;115(23):2948–56.
- 67. Aikawa M, Libby P. The vulnerable atherosclerotic plaque: pathogenesis and therapeutic approach. Cardiovasc Pathol. 2004 Jun;13(3):125–38.
- Vieceli Dalla Sega F, Fortini F, Aquila G, Campo G, Vaccarezza M, Rizzo P. Notch Signaling Regulates Immune Responses in Atherosclerosis. Front Immunol. 2019; 10:1130.