

The Morphometric Analysis of the Effects of Hydrogen Peroxide on the Off Springs of Helix Lymnaea Model

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ABSTRACT

Background: Hydrogen peroxide free radical is a potent mutagen and teratogen that can obviously affects the color of the offspring's of the land snail Helix Lymnaea.

Aim: To assess the morphometric changes of the offspring's of the land snail Helix Lymnaea after parents exposure for H2O2 for three years.

Materials and Methods: A twenty four Helix Lymnaea snails were randomly divided into two groups (12 in each). Group A is the control whereas group B is the H2O2 test group. All were adults (4-6 weeks age) of the same size 5-7 mm length, all were hermaphrodites. The two groups were followed for three years for ten generations for different morphometric parameters

Results: There was a significant reduction in the movement score in the group treated by H2O2 in comparison with the control group 25% vs 8.3 % respectively p = 3.8, chi square = 0.8. The eating activity of the offspring's had a minute change noticed in between the two groups 95% in H2O2 exposed vs 92% in the control group. The morphometric outcomes mainly the albino offsprings were none in the

control group that showed no teratogenicity in comparison with 60% in H2O2 group, p < 0.05. The rate of successful eggs hatching was significantly differed between the control and the H2O2 groups, p = 0.017. There was also an obvious difference in the mortality rate between the control and H2O2 group, however this difference was not significant p = 0.2.

Conclusion: There was a significant teratogenic effect of H2O2 on the offsprings of the snail Helix Lymnaea. This effects may extend to include different free radical producing substances.

Key words: Helix Lymnaea, H2O2, morphometry, reproductive rate, teratogenicity.

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INTRODUCTION

Mutagenesis, teratogenesis and carcinogenesis are the pathological processes that alter the genome wild integrity and hence initiate a real health challenges not only for human rather for all of the living species and the ecosystem (Tchounwou, et al, 2012).

Environmental teratogens are so many that originate from different sources in our ecosystem such as chemicals, radiation and viral origins (Parsa, 2012). Of the most potent chemical mutagen is the free radicals (Phaniendra, et al, 2015) of which oxygen radical species H2O2 are the most important (Wen, et al. 2013).

Many byproducts from household and factories are toxic to the biosystems like marshes, runnels and soil (Lobo, et al. 2010). Mutagenic byproduct for any biosystem is a potential toxin and mutagen for the remaining biosystems including the human being (DeVito, et al. 2017). H2O2 sharing the same mechanism of mutagenesis and teratogenesis with any reactive radical in that they react directly with nucleotides or facilitate their epigenetic reactions like methylation, oxidation or acetylation (Afanas'ev , 2015). The net result is an altered DNA base pairing and epigenetic structure (Afanas'ev, 2013). This mechanism is the commencement step of cell transformation and teratogenesis (Afanas'ev , 2010).

Different models are adopted to predict the level by which a test chemical exerts morphometric level of

teratogenicity. Common in vitro models involve Ames test in typhimurium species, Escherichia mutagenic tests, karyotyping, cell culture models (Samiei, et al. ,2015) whereas mice, rat, rabbits and monkeys are common animal models (Badyal & Desai, 2014). Different snail species are suitable to be a reliable model of environmental pollution and its effects on offsprings. One of these helices is the Helix Lymnaea (Regoli, et al.,2006). This helix type has favourable features such as its high productive rate. Its continuous reproduction, high amount of offsprings per each generation, short period of maturation of offsprings to adults, its aqueous watery environment makes it accessible with test chemicals and easily monitored by morphometric image analyzer. Moreover Helix Lymnaea can sustain extreme environments which enables a prolonged years of monitoring (Regoli , et al. 2006; Abdel-Halim, et al. 2013& Gomot-de, 2002). The research hypotgesis of this current study which assume a potential effects of environmental oxygen radicals on the morphology of living organisms generations.

STUDY OBJECTIVES

To assess the morphometric outcomes for the offsprings of Helix Lymnaea after 3 years of exposure to environmental H2O2.

MATERIALS AND METHODS

Table 1: Materials used, vendors, dosage form and country of source

Materials	Vendor	Dosage form	Country	ISO
H2O2	Fisher	500 ml liquid	Germany	Yes
Distilled water	Local distilator	5 L	Local	-
Soil	Natural runnels	Semisolid	Local	-

Runnels water	Natural runnels	Liquid	Local	-
Formalin	Fisher	1 L Liquid	Germany	Iso

Table 2: Instruments and tools used, their company, models, and country of origin.

Instruments and lab tools	Company model	Country	Iso/ properties
Camera	Zennet	Russia	Yes
Light microscope	Ziets	Germany	Yes
Image processor for morphometry on PC	Matlab	-	-
Glass containers	Bayer	Germany	1 L / 3 containers
Sensitive balance	-	Germany	3 decimals
Volumetric flasks	Bayer	Germany	500 ml
Dropper	-	Germany	5 ml
Micropipet	Eppendorph	Germany	2 microml

Study design

The current research designed an experimental assessment of the morphological analysis of a three years H₂O₂ exposure in Helix Lymnaea model.

Twenty four Helix Lymnaea snails were divided into two groups (12 in each). Group A is the control whereas group B is the H₂O₂ test group.

All snails were adults (4-6 weeks age) of the same size 5-7 mm length. The sex was hermaphroditism. The study was conducted in a private lab in Baghdad for three years.



Figure 1: Shows the species used of the snail Helix Lymnaea. It was aquatic and hermaphrodite.



Figure 2: The culture prepared for breeding Helix Lymnaea. It reveals the aggregates of eggs within a shell of gel. It was used to count eggs laying and hatching rate as an indicator of the reproductive activity.

Groups division

The 24 Helix Lymnaea were divided into the following:

1. The control group (N = 12) were given runnels soil (5 grams every day) within a 1 liter dandom runnel water. The culture was washed and replaced with the next 5 grams soil every day.

2. H₂O₂ group (N = 12) were given 5% H₂O₂ in their culture. Otherwise similar preparation to the control group.

Sample selection

The snail sample selection:

A random selection of snails was followed from different areas of the runnels

The soil and runnels water selection

The random way of selecting a sample was also followed.

Parameters to be monitored

Monitoring and record registry were done at rate demonstrated per each parameter measured and along ten generations throughout 3 years of follow up. The snails reproductive activity was diminished during fall and winter and hence the monitoring was postponed until the next spring when their activity was recruited.

1. Number of active, inert and died snail every week along the course

2. Soil eating activity score (as indicated in waste fraction out of 5 grams of soil per day. This parameter can be indicated by image processing software)

3. Active movement score (1 = low, 2 = moderate, 3 = high activity)

4. Weekly culture image for morphometric assessment

5. Reproductive Rate (Number of hatching egg per month and number of egg laying per month)

7. Ratio of monthly successful egg hatching

8. Death rate monthly

RESULTS

Table 3: The number of active and inert snails (active is a continuously moving, reproducing and eating snail).

Groups	Active number	Inactive number	Statistics
Control	12	1	SD = 1 p = 0.38 Chi square = 0.8
H2O2	10	3	

Table 4: The soil eating activity (fraction of waste in relation to the 5 grams of soil introduced daily to the culture).

Groups	Eating activity (waste/food percentage)	Statistics
Control	95	SD = +/- 6
H2O2	92	SD = +/- 5

Culture morphometric analysis

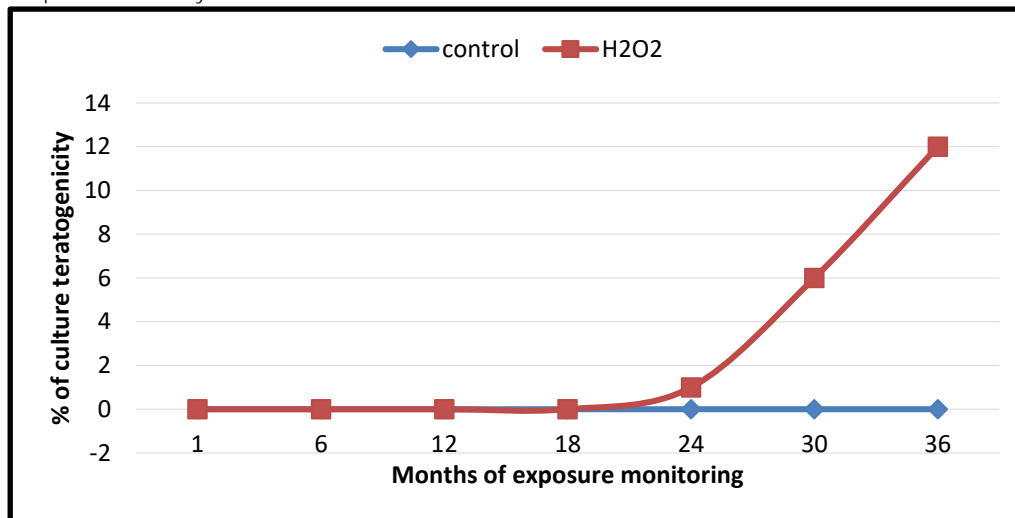


Figure 3: The percentage of teratogenicity recorded for the offsprings of Helix Lymnaea in form of albino generations.

REPRODUCTIVE RATE

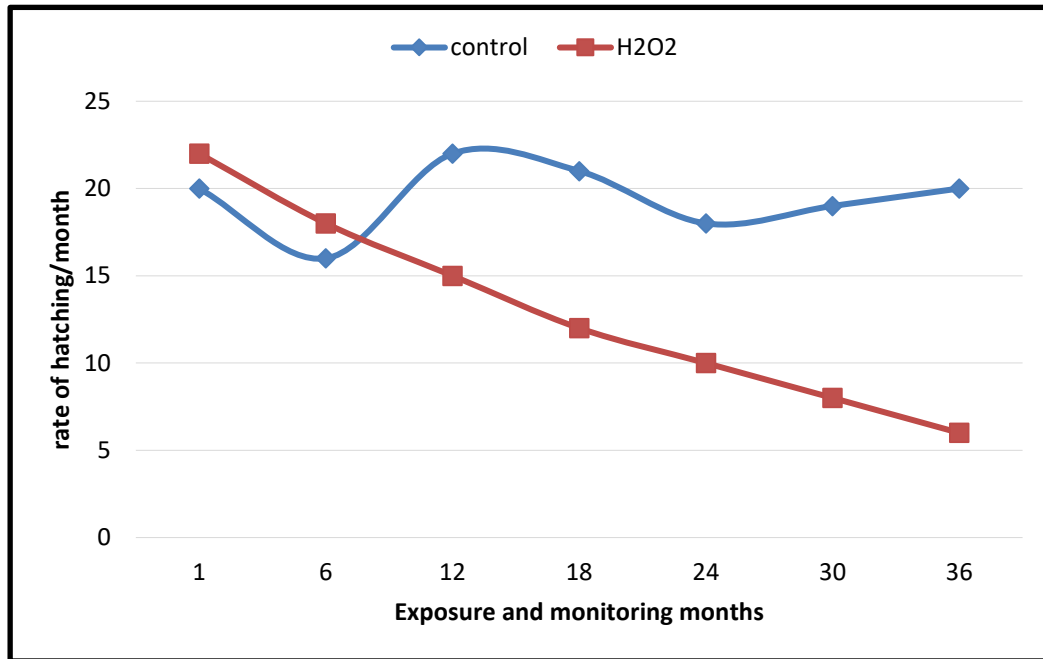


Figure 4: Changes in eggs hatching rate as a response to exposure to H2O2 throughout 36 months.

MORTALITY PERCENTAGES

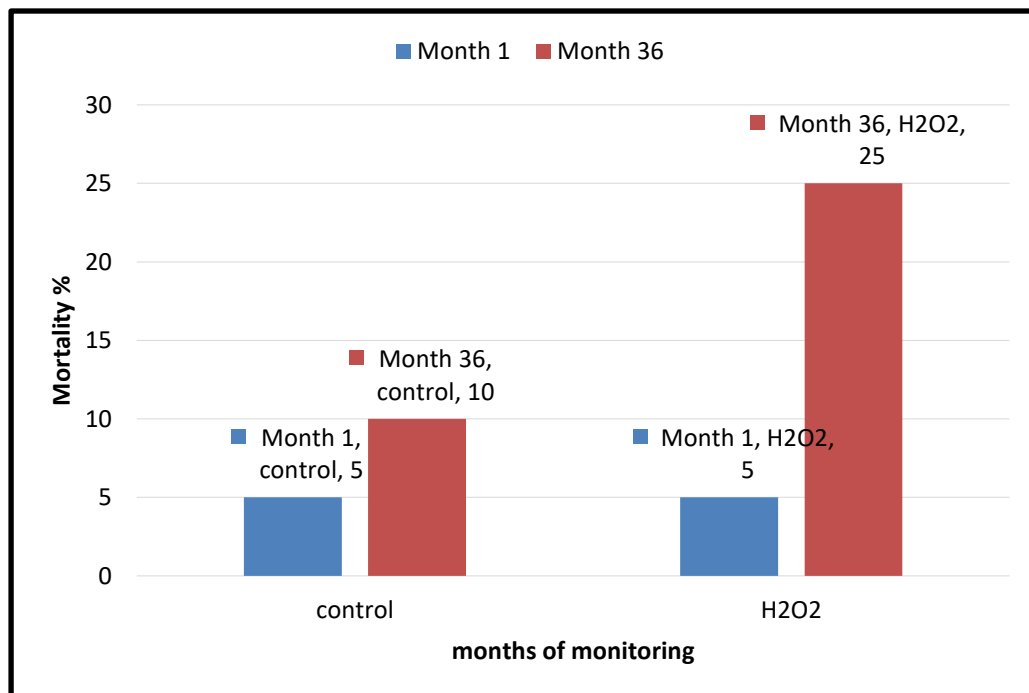


Figure 5: The mortality rate in response to a 36 months exposure to H2O2 as compared with the control group.

DISCUSSION

Any model of assessing the impact of a substance on offsprings and teratogenicity has to be characterized by a high reproductive rate (Forray & Foster, 2015) in form of high egg laying rate and high successful rate of hatching (Janssen & Baur, 2015). Moreover, the aqueous media animals are more prone for the teratogenic effects of environmental xenobiotics (Brent, 2004) since water and organic soluble substances are readily diffused into the organs (Brent, 2004). The model of Helix Lymnaea is adequately fulfilling these criteria.

The results showed that there was a significant reduction in the movement score in the group treated by H2O2 in comparison with the control group 25% vs 8.3 % respectively $p = 3.8$, $\chi^2 = 0.8$. This reduction in the movement of the Helix may be attributed to the direct impact of the free radicals on the muscles (Ji et al, 2009) or it may be due to the mutagenic effect of H2O2 on different cell components genes (Netto & Antunes, 2016). Many other environmental factors like pesticides and organophosphorous may also contribute to change in aquatic animals movements (Jayaraj et al 2016 & Katagi, 2010).

Another property had been assessed as a general parameter for the impact of H₂O₂ on the generations of the Helix Lymnaea was the eating activity of the offsprings as indicated by the amount of waste produced over the amount of food introduced into the culture. There was a minute change noticed in between the two groups 95% in H₂O₂ exposed vs 92% in the control group.

The morphometric outcomes mainly referred to as the albino offsprings since those were the most prominent teratogenic effect of mutagenic mechanism since the exposure was to the parent snails. The control group showed no change in teratogenicity in comparison with 60% in H₂O₂ group, $p < 0.05$. Researches had verified color change in different species induced by several xenobiotic agents (Raunio et al. 2015; Moon et al. 2006).

The rate of successful eggs hatching was another important parameter measured. This indicate the healthy sperms, eggs and zygotes that were unaffected by the xenobiotic. There was a significant difference between the control and the H₂O₂ groups, $p = 0.017$. Studies that verified the influence of free radicals on the rate of successful egg hatching for different species include (Wong et al. 2008; Wong et al. 2004 & Rehkopf et al. 2017).

There was obvious difference in the mortality rate between the control and H₂O₂ group, however this difference was not significant $p = 0.2$. There are many environmental factors can cause an increase in mortality rate like pH, temperature, and chemical contamination (Bighiu et al. 2017). Of these chemical contaminants are the bisphenols (Corrales et al. 2015) and metals (Notten et al. 2005).

CONCLUSION

There was a significant teratogenic effect of H₂O₂ on the offsprings of the snail Helix Lymnaea. This effect may extend to include different free radical producing substances.

RECOMMENDATION

We recommend to conduct researches of free radicals producing drugs in similar models like tissue cultures and amphibians to assess the genetic and epigenetic alterations as a result of such exposure.

CONFLICT OF INTEREST

None of the authors have any conflicts of interest relevant to this research subject.

ETHICAL CLEARANCE

The study was conducted in accordance with ethical principles that have their origin in the Declaration of Helsinki. The study protocol, care of animals and subject information were reviewed and approved by a local Ethics Committee.

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None.

REFERENCES

1. Tchounwou PB, Yedjou CG, Patlolla AK, Sutton DJ.(2012) Heavy metal toxicity and the environment. *Exp Suppl.*:101:133-64.
2. Parsa N. (2012) Environmental factors inducing human cancers. *Iran J Public Health.*:41(11):1-9
3. Phaniendra A, Jestadi DB, Periyasamy L.(2015) Free radicals: properties, sources, targets, and their implication in various diseases. *Indian J Clin Biochem.*:30(1):11-26
4. Wen X, Wu J, Wang F, Liu B, Huang C, Wei Y.(2013) Deconvoluting the role of reactive oxygen species and autophagy in human diseases. *Free Radic Biol Med.*:65:402-410
5. Lobo V, Patil A, Phatak A, Chandra N. (2010) Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacogn Rev.*:4(8):118-26
6. DeVito S, Woodrick J, Song L, Roy R.(2017) Mutagenic potential of hypoxanthine in live human cells. *Mutat Res.* :803-805:9-16
7. Afanas'ev I.(2015) Mechanisms of superoxide signaling in epigenetic processes: relation to aging and cancer. *Aging Dis.* 1;6(3):216-27
8. Afanas'ev I.(2013) New nucleophilic mechanisms of ros-dependent epigenetic modifications: comparison of aging and cancer. *Aging Dis.* 21;5(1):52-62
9. Afanas'ev I. (2010) Signaling and Damaging Functions of Free Radicals in Aging-Free Radical Theory, Hormesis, and TOR. *Aging Dis.*:1(2):75-88
10. Samiei M, Asgary S, Farajzadeh M, Bargahi N, Abdolrahimi M, Kananizadeh U, et al.(2015) Investigating the mutagenic effects of three commonly used pulpotomy agents using the ames test. *Adv Pharm Bull.*:5(1):121-5.
11. Badyal DK, Desai C (2014). Animal use in pharmacology education and research: the changing scenario. *Indian J Pharmacol.*:46(3):257-65.
12. Regoli F, Gorbi S, Fattorini D, Tedesco S, Notti A, Machella N, et al.(2006) Use of the land snail Helix aspersa as sentinel organism for monitoring ecotoxicologic effects of urban pollution: an integrated approach. *Environ Health Perspect.*:114(1):63-9.
13. Gomot de Vaufleury A, Pihan F.(. 2000) Growing snails used as sentinels to evaluate terrestrial environment contamination by trace elements. *Chemosphere.*:40(3):275-84.
14. Abdel-Halim KY, Abo El-Saad AM, Talha MM, Hussein AA, Bakry NM.(2013) Oxidative stress on land snail Helix aspersa as a sentinel organism for ecotoxicological effects of urban pollution with heavy metals. *Chemosphere.*:93(6):1131-8.
15. Gomot-de VA, Pihan F.(2002) Methods for toxicity assessment of contaminated soil by oral or dermal uptake in land snails: metal bioavailability and bioaccumulation. *Environ Toxicol Chem.*:21(4):820-7.
16. Forray A, Foster D.(2015) Substance Use in the Perinatal Period. *Curr Psychiatry Rep.*:17(11):91
17. Janssen R, Baur B.(2015) Seasonal effects on egg production and level of paternity in a natural population of a simultaneous hermaphrodite snail. *Ecol Evol.*:5(14):2916-28
18. Brent RL. (2004) Utilization of animal studies to determine the effects and human risks of environmental toxicants (drugs, chemicals, and physical agents). *Pediatrics.*:113(4 Suppl):984-95.

19. Ji LL, Gomez-Cabrera MC, Vina J. (2009) Role of free radicals and antioxidant signaling in skeletal muscle health and pathology. *Infect Disord Drug Targets*.;9(4):428-44
20. Netto LE, Antunes F (2016). The Roles of Peroxiredoxin and Thioredoxin in Hydrogen Peroxide Sensing and in Signal Transduction. *Mol Cells*.;39(1):65-71
21. Jayaraj R, Megha P, Sreedev P (2016). Organochlorine pesticides, their toxic effects on living organisms and their fate in the environment. *Interdiscip Toxicol*.;9(3-4):90-100.
22. Katagi T (2010). Bioconcentration, bioaccumulation, and metabolism of pesticides in aquatic organisms. *Rev Environ Contam Toxicol*.;204:1-132.
23. Raunio H, Kuusisto M, Juvonen RO, Pentikäinen OT (2015). Modeling of interactions between xenobiotics and cytochrome P450 (CYP) enzymes. *Front Pharmacol*. 12;6:123
24. Moon YJ, Wang X, Morris ME (2006). Dietary flavonoids: effects on xenobiotic and carcinogen metabolism. *oxicol In Vitro. r*;20(2):187-210
25. Wong JL, Wessel GM (2008). Free-radical crosslinking of specific proteins alters the function of the egg extracellular matrix at fertilization. *Development*.;135(3):431-40
26. Wong JL, Créton R, Wessel GM (2004) . The oxidative burst at fertilization is dependent upon activation of the dual oxidase Udx1. *Dev Cell*.;7(6):801-14.
27. Rehkopf AC, Byrd JA, Coufal CD, Duong T (2017). Advanced Oxidation Process sanitization of hatching eggs reduces Salmonella in broiler chicks. *Poult Sci*. 1;96(10):3709-3716.
28. Bighiu MA, Gorokhova E, Carney Almroth B, Eriksson Wiklund AK (2017). Metal contamination in harbours impacts life-history traits and metallothionein levels in snails. *PLoS One*. 3;12(7):e0180157
29. Corrales J, Kristofco LA, Steele WB, Yates BS, Breed CS, Williams ES, Brooks BW (2015). Global Assessment of Bisphenol A in the Environment: Review and Analysis of Its Occurrence and Bioaccumulation. *Dose Response*. 29;13(3):1559325815598308.
30. Notten MJ, Oosthoek AJ, Rozema J, Aerts R (2005). Heavy metal concentrations in a soil-plant-snail food chain along a terrestrial soil pollution gradient. *Environ Pollut*.;138(1):178-90.